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Antibacterial effect of pepper and cumin extracts on some pathogens contaminating chicken meat

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ABSTRACT

Prevention of food poisoning bacteria is usually achieved by using of some chemical preservatives which have public hazards coming through the improper use of it including presence of chemical residues in food, and acquisition of microbial resistance. Based on such safety concerns, the public need to replace it with more potentially effective, healthier natural alternative preservatives are increased. In the present study, two plant extracts, represented by black pepper and cumin extracts were examined to inhibit the growth of foodborne *Staphylococcus aureus*, *Escherichia coli* and *Salmonella Typhi* strains that were artificially inoculated into minced chicken meat samples. Results revealed that the used pepper and cumin extracts significantly reduced the count of tested strains by 10^2 CFU/g with reduction percent exceeded 99.0%, which proved that the used herbal extracts were potentially effective with variable efficiency against the tested bacterial strains; so, it can be used as natural alternative preservatives to control food poisoning diseases and preserve food stuff avoiding health hazards of chemically antimicrobial agent applications.

1. INTRODUCTION

Thousands of human deaths are reported yearly due to foodborne illness in developing countries Sapkota et al. (2012). Most of food related illness is associated with bacteria and/or its toxins contaminating foods especially *E. coli* and *S. Typhi* as Gram negative bacteria (Pandey and Singh, 2011). Moreover, *Staphylococcus aureus*, as Gram positive bacteria, was reported as the 3rd foodborne pathogen associated with food poisoning which essentially referred to its wide variety of enterotoxins production (Normanno et al., 2007).

Control of foodborne pathogens is commercially performed by addition of chemical preservatives (Shan et al., 2007). As they have antibacterial activity, many researches were conducted to replace them by natural preservatives to avoid their hazards due to their repeated applications such as accumulation of chemical residues, microbial resistance, and side effects on human health (Nasar-Abbas and Kadir, 2004; Bialonska et al., 2010).

Different plants are known of their maintenance ability of good human health since long time ago. In addition, the interest in natural products as food preservatives has greatly increased because of development of multiple antibiotic resistance pathogens that increases interest in the use of herbal extracts as antimicrobial agents (Abdallah and Koko, 2017).

Natural herbal extracts of medicinal purposes contain high biologically active compounds which have great bacterial inhibition properties. Natural plants spices and extracts of

various plant parts have been used widely, especially in traditional cultures, as natural antimicrobials, antioxidants, and flavoring agents (George et al., 2009).

Black pepper (*Piper nigrum*) is one of the most public flavoring spices over the world. It is known as the king of flavors. In addition to its flavoring properties, it has many bio-active effects on the human body, such as its perceptible antibacterial action (Abdallah and Abdalla, 2018).

Cumin (*Cuminum cyminum*) is one of these herbs; it is an herbaceous annual plant used as a relish ingredient in many foods processing not only for its seasoning and flavoring effect, but also as natural preservative due to its inhibitory effect against many bacteria (Bahraminejad et al., 2010).

Both black pepper and cumin characterized by medicinal and health benefits; where cumin is used to reduce GIT inflammation, carminative effect, and suppress muscle spasms; moreover, it is also used in improving ingestion, jaundice, diarrhea and flatulence (Eikani et al., 2007). In addition, black pepper and its bioactive extract especially the chemical piperine that has numerous physical and antimicrobial actions. Singletary (2010) documented that black pepper may have health benefits, particularly in enhancing the function of GIT, nervous system benefits, and may influence body energy; also the author suggested that black pepper contains antioxidant constituents and possesses anti-inflammatory and antimicrobial properties.

Therefore, this study aimed to investigate the antibacterial effect of pepper and cumin extracts against *S. aureus*, *E. coli*, and *S. Typhi* in the artificially inoculated minced chicken meat samples.

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2. MATERIAL AND METHODS

2.1. Collection of samples

2.1.1. Meat samples

About 2000 g of fresh chicken meat samples were collected from poultry butchery located in Menoufiya Governorate, Egypt. Samples were minced and divided into equal portions (200g) and kept in separate plastic bags and refrigerated at 4 °C.

2.1.2. Herbal extracts

Black pepper and cumin extracts were obtained from Animal Health Research Institute, Shebin Elkom Lab.

2.1.3. Bacterial strains

Staphylococcus aureus, *Escherichia coli* and *Salmonella Typhi* strains were obtained from the Central Food Quality Lab., Faculty of Veterinary Medicine, Benha University.

2.2. Experimental application

2.2.1. Minced meat samples

After Ultraviolet sterilization, minced meat samples were divided into 4 groups, 1st untreated, 3 treated groups (200 g for each) and classified into:

2.2.2. Black pepper extract (Kaur et al., 2017)

- 1st group: control group kept in 4 °C without any additives.
- 2nd group: inoculated with of *S. Typhi* 10⁵ (CFU/g) + pepper extract (150 mg/ml).
- 3rd group: inoculated with of *S. aureus* (10⁶ CFU/g) + pepper extract (150 mg/ml).
- 4th group: inoculated with of *E. coli* (10⁴ CFU/g) + pepper extract (150 mg/ml).

2.2.3. Cumin extract

- 1st group: control group kept in 4 °C without any additives.
- 2nd group: inoculated with of *S. Typhi* (10⁵ CFU/g) + cumin extract (150 mg/ml) (Sheikh et al., 2010).
- 3rd group: inoculated with of *S. aureus* (10⁶ CFU/g) + cumin extract (150 mg/ml) (Mostafa et al., 2018).
- 4th group: inoculated with of *E. coli* (10⁴ CFU/g) + cumin extract (150 mg/ml) (Sheikh et al., 2010).

Both untreated and treated samples were packed in a separate plastic bags, stored at (4±1°C), and then examined bacteriologically after (2 hrs) as zero time and at predetermined interval (24 hrs) till decomposition of meat has occurred organoleptically after the 7th day of the experiment. The experiment was conducted in triplicate. Colonies of the examined samples in each group were counted and recorded after the preparation of samples according to APHA (2013) to obtain ten-fold serial dilution through mixing of 25 grams of each sample with 225 ml of 0.1% peptone water, the contents were homogenized at stomacher at 450 to 640 strokes/min for 2 minutes; 1ml of the homogenization was transferred into separate tubes containing 9 ml of sterile peptone water 0.1%, from which ten-fold serial dilutions were prepared.

2.3. *S. aureus* count was done according to ISO 6888-1 (1999), A1 (2003).

2.4. *E. coli* count was performed according to ISO 16649-2 (2001).

2.5. *S. Typhi* count was performed according to ISO 6579-1 (2017).

2.6. Statistical analysis:

The obtained results were statistically evaluated by application of Analysis of Variance (ANOVA) test according to Feldman et al. (2003).

3. RESULTS

Application of black pepper and cumin extracts as natural herbal preservative to control *S. aureus*, *E. coli* and *S. Typhi* in minced chicken meat samples showed significant inhibitory effect with reduction rate exceeded 99.0%.

Regarding the effect of herbal extracts on artificially inoculated *S. aureus* minced chicken meat sample, table (1) revealed that the mean value of examined strain count at zero time was $2.3 \times 10^6 \pm 5.7 \times 10^4$ CFU/g in control and treated groups. After seven days, *S. aureus* counts showed significant reduction with mean value of $5.5 \times 10^4 \pm 3.0 \times 10^3$ and $6.5 \times 10^4 \pm 1.8 \times 10^3$ (CFU/g) and reduction rate of 99.9% for both pepper and cumin treated groups, respectively. While *S. aureus* counts was $1.9 \times 10^8 \pm 2.3 \times 10^7$ CFU/g in the control group. Significant differences between the bacterial count/group were noticed at P 0.05.

The inhibitory effect of tested herbal extracts on artificially inoculated *E. coli* strain into minced chicken meat sample was estimated. Results tabulated in table (2) showed that the mean count of *E. coli* at zero time was $2.2 \times 10^4 \pm 8.8 \times 10^4$ in control and treated groups. The mean value of *E. coli* after the 7th day of experiment in control group was $3.5 \times 10^6 \pm 5.0 \times 10^5$ CFU/g. While in treated groups, *E. coli* showed significant reduction with mean value of $4.7 \times 10^2 \pm 2.4 \times 10$ and $6.7 \times 10^2 \pm 1.8 \times 10$ CFU/g with reduction rate of 99.98% for both pepper and cumin treated groups, respectively. Significant differences between the bacterial counts were at P 0.05.

Table (3) presented the inhibitory effect of cumin and pepper extracts on artificially inoculated *S. Typhi* minced chicken meat sample. The mean count at zero time was $2.2 \times 10^5 \pm 1.2 \times 10^4$ CFU/g in control and treated groups. At 7th day of the experiment, the mean value of *S. Typhi* was $3.8 \times 10^7 \pm 1.7 \times 10^6$ CFU/g in control group, but it was $7.5 \times 10^2 \pm 0.8 \times 10$ and $8.1 \times 10^2 \pm 1.4 \times 10$ CFU/g with reduction percent of 99.9% for both pepper and cumin treated groups, respectively. Significant differences between the bacterial counts were at P 0.05.

4. DISCUSSION

Herbal spices have been used for centuries by many ancient cultures for improving the flavor and aroma of foods. In addition, they have been used as food preservative due to its antimicrobials properties (Pan et al., 2008). Referring to the global foodborne illness reports, bacterial food poisoning represents two third of foodborne disease outbreaks (Sodha et al., 2009). The main microorganism related to manipulation practices, *Staphylococcus* spp., *E. coli* and *Salmonella* spp. (Lukinmaa et al., 2004).

Referring to the present obtained results, usage of black pepper and cumin extracts showed promising inhibitory effects against foodborne tested *S. aureus*, *E. coli* and *S. Typhi*. In line with many previous studies that recorded great antibacterial effects against many foodborne bacteria. Comparing with the previous studies, the inhibitory effect of black pepper effect could be compared with the records of Abdel Gadir et al. (2007) revealed inhibitory effect of

methanol extract of pepper against *E. coli* and *S. Typhi*, while had no activity against *S. aureus*. Moreover, petroleum ether extract had no activity against all tested species, while ethanolic extract revealed lower inhibitory activity. Sulieman and Allaahmed, (2012) reported a dramatic decrease in *S. aureus* count to 2.1×10^4 CFU/g after treatment with 5% pepper for 96 hrs. Rani et al. (2013) documented that the inhibitory effect of black pepper extract was at maximum against Gram positive bacteria *S. aureus* and but at minimum against Gram negative bacteria *E. coli*. Nagy et al. (2015) recorded inhibitory effect of methanol extract of pepper against *E. coli* and *S. aureus*. Zou et al. (2015) showed inhibitory effect of chloroform extract of pepper against *E. coli* and *S. aureus*. Ibrahim et al. (2016) declared bactericidal effect of methanol extract of pepper against *E. coli* and *S. Typhi*, while lesser inhibition against *S. aureus*. Kaho et al. (2019) recorded low inhibitory effect of pepper against *E. coli*.

Regarding to the present inhibitory effect of cumin extract, it could be compared with the records of Dua et al. (2013), who mentioned that cumin extract was effective against both Gram-positive and -negative bacteria which was

referred to causing of cell membrane damage and release of the intracellular nucleotides and proteinaceous material. Mostafa et al. (2018), reported that the effect of cumin extract (5-10 mg/ml) was only effective against *S. aureus*, while it was not effective against *E. coli* and *S. Typhi*. Abdul Jabbar (2013) recorded that cumin oil exhibited a strong antibacterial activity against *E. coli*, *S. aureus* isolated from food samples, Baljeet et al. (2015) reported lesser inhibition effect of cumin against *S. Typhi*.

According to the aforementioned studies, different black pepper and cumin extracts showed significant inhibitory effects against varied gram-positive and gram-negative bacteria, although some studies revealed that some extracts had no antibacterial activity, which could be attributed to differences in plant varieties, microbiological methods, solvents used and tested microorganisms. In general, the majority of these studies suggested that the black pepper and cumin extracts could be a potential candidate for developing new food preservative against wide ranges of pathogenic bacteria either food borne, food spoilage or clinical isolates.

Table 1 Statistical analytical results of the effect of some herbal extracts on artificially inoculated *Staphylococcus aureus* into mined chicken meat samples during different period of cold storage (4 °C).

Time	Control	Group A (pepper)		Group B (cumin)	
	Mean ± S.E.	Mean ± S.E.	R%	Mean ± S.E.	R%
Zero time	$2.3 \times 10^6 \pm 5.7 \times 10^{4d}$	$2.3 \times 10^6 \pm 5.7 \times 10^{4a}$	-	$2.3 \times 10^6 \pm 5.7 \times 10^{4a}$	-
2 nd day	$4.6 \times 10^6 \pm 1.5 \times 10^{5cd}$	$1.3 \times 10^6 \pm 5.7 \times 10^{4b}$	71.74	$1.6 \times 10^6 \pm 8.8 \times 10^{4b}$	65.22
3 rd day	$8.3 \times 10^6 \pm 1.1 \times 10^{5cd}$	$9.2 \times 10^5 \pm 8.8 \times 10^{3c}$	88.92	$9.8 \times 10^5 \pm 5.7 \times 10^{3c}$	88.19
4 th day	$9.5 \times 10^6 \pm 1.2 \times 10^{5cd}$	$6.3 \times 10^5 \pm 1.1 \times 10^{4d}$	93.37	$6.7 \times 10^5 \pm 8.8 \times 10^{3d}$	92.94
5 th day	$3.3 \times 10^7 \pm 1.1 \times 10^{6c}$	$2.3 \times 10^5 \pm 1.4 \times 10^{4e}$	99.30	$3.7 \times 10^5 \pm 1.1 \times 10^{4e}$	98.87
6 th day	$6.2 \times 10^7 \pm 8.8 \times 10^{5b}$	$8.2 \times 10^4 \pm 5.7 \times 10^{2f}$	99.86	$9.6 \times 10^4 \pm 2.5 \times 10^{3f}$	99.84
7 th day	$1.9 \times 10^8 \pm 2.3 \times 10^{7a}$	$5.5 \times 10^4 \pm 3.0 \times 10^{3f}$	99.99	$6.5 \times 10^4 \pm 1.8 \times 10^{3f}$	99.97

The values represent Mean ± SE of three experiments. Means within a column followed by different letters are highly significantly different ($P < 0.05$). Zero time: 2h after inoculation. R%: Reduction percent.

Table 2 Statistical analytical results of the effect of some herbal extracts on artificially inoculated *Escherichia coli* into mined chicken meat samples during different period of cold storage (4 °C).

Time	Control	Group A (pepper)		Group B (cumin)	
	Mean ± S.E.	Mean ± S.E.	R%	Mean ± S.E.	R%
Zero time	$2.2 \times 10^4 \pm 8.8 \times 10^2$	$2.2 \times 10^4 \pm 8.8 \times 10^4$	-	$2.2 \times 10^4 \pm 8.8 \times 10^4$	-
2 nd day	$5.4 \times 10^4 \pm 8.8 \times 10^2$	$1.2 \times 10^{4b} \pm 1.2 \times 10^3$	75.92	$1.7 \times 10^{4b} \pm 1.2 \times 10^3$	68.52
3 rd day	$8.3 \times 10^4 \pm 8.8 \times 10^2$	$7.4 \times 10^{3c} \pm 1.2 \times 10^2$	91.08	$8.5 \times 10^{3c} \pm 1.7 \times 10^2$	89.76
4 th day	$5.2 \times 10^{5bc} \pm 5.7 \times 10^3$	$5.2 \times 10^{3d} \pm 8.8 \times 10$	99.00	$6.8 \times 10^{3d} \pm 5.7 \times 10$	98.69
5 th day	$7.7 \times 10^{5b} \pm 8.8 \times 10^3$	$1.4 \times 10^{3e} \pm 1.2 \times 10^2$	99.81	$2.4 \times 10^{3e} \pm 2.3 \times 10^2$	99.68
6 th day	$9.6 \times 10^{5b} \pm 1.8 \times 10^4$	$8.3 \times 10^{2e} \pm 0.5 \times 10$	99.91	$9.4 \times 10^{2e} \pm 1.1 \times 10$	99.90
7 th day	$3.5 \times 10^{6a} \pm 5.0 \times 10^5$	$4.7 \times 10^{2b} \pm 2.4 \times 10$	99.98	$6.7 \times 10^{2b} \pm 1.8 \times 10$	99.98

The values represent Mean ± SE of three experiments. Means within a column followed by different letters are highly significantly different ($P < 0.05$). Zero time: 2h after inoculation. R%: Reduction percent.

Table 3 Statistical analytical results of the effect of some herbal extracts on artificially inoculated *Salmonella typhi* into mined chicken meat samples during different period of cold storage (4 °C).

Time	Control	Group A (pepper)		Group B (cumin)	
	Mean ± S.E.	Mean ± S.E.	R%	Mean ± S.E.	R%
Zero time	$2.2 \times 10^5 \pm 1.2 \times 10^{4e}$	$2.2 \times 10^5 \pm 1.2 \times 10^{4a}$	-	$2.2 \times 10^5 \pm 1.2 \times 10^{4a}$	-
2 nd day	$4.5 \times 10^5 \pm 2.4 \times 10^{4de}$	$1.5 \times 10^5 \pm 2.8 \times 10^{4b}$	66.6	$1.7 \times 10^5 \pm 1.2 \times 10^{4b}$	62.2
3 rd day	$8.5 \times 10^5 \pm 1.7 \times 10^{4de}$	$8.1 \times 10^4 \pm 2.0 \times 10^{3c}$	90.5	$8.6 \times 10^4 \pm 1.4 \times 10^{3c}$	89.9
4 th day	$2.5 \times 10^6 \pm 1.7 \times 10^{5d}$	$5.6 \times 10^4 \pm 1.4 \times 10^{3cd}$	97.8	$6.3 \times 10^4 \pm 1.4 \times 10^{3d}$	97.5
5 th day	$5.5 \times 10^6 \pm 1.7 \times 10^{5c}$	$2.2 \times 10^4 \pm 8.8 \times 10^{3de}$	99.6	$3.2 \times 10^4 \pm 1.2 \times 10^{3e}$	99.4
6 th day	$9.6 \times 10^6 \pm 1.4 \times 10^{5b}$	$1.5 \times 10^3 \pm 1.2 \times 10^{2c}$	99.9	$3.7 \times 10^3 \pm 6.6 \times 10^{2f}$	99.9
7 th day	$3.8 \times 10^7 \pm 1.7 \times 10^{6a}$	$7.5 \times 10^2 \pm 0.8 \times 10^0$	99.9	$8.1 \times 10^2 \pm 1.4 \times 10^0$	99.9

The values represent Mean ± SE of three experiments. Means within a column followed by different letters are highly significantly different ($P < 0.05$). Zero time: 2h after inoculation. R%: Reduction percent.

5. CONCLUSION

From the present results, it can be concluded that herbal extracts of black pepper and cumin had potential inhibitory effects against foodborne bacterial contamination giving promising trend in replacing chemical preservatives with natural safe herbal extracts.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

6. REFERENCES

- Abdallah, E.M. and Abdalla, W.E. 2018. Black pepper fruit (*Piper nigrum* L.) as antibacterial agent: A mini-review. *J. Bacteriology and Mycology*, 6(2): 141-145.
- Abdallah, E.M. and Koko, W.S. 2017. Medicinal plants of antimicrobial and immunomodulating properties. In: *Antimicrobial research: Novel bio-knowledge and educational programs*. Méndez-Vilas, A. (ed.). Formatex Research Center, Spain. Pp. 127-139.
- Abdel Gadir, W.S., Mohamed, F., Bakhiet, A. 2007. Antibacterial activity of *Tamarindus indica* fruit and *Piper nigrum* seed. *Research Journal of Microbiology*, 2(11): 824-830.
- Abdul Jabbar, R.A. 2013. Chemical analysis and antimicrobial activity of Cumin seeds extracted oil against some bacterial and fungal isolates *Journal of Thi-Qar Science's*, 3(4): 65-73.
- APHA (American Public Health Association) 2013. *Compendium of methods for the microbiological examination of food*. T. Matthew Taylor, John N. Sofos, Peter Bodnaruk, and Gary R. Acuff (Eds.), 4th Ed., Chapter. 2, Washington DC., USA.
- Bahraminejad, A., Mohammed-Nejad, G., Abdulkhadir, M. 2010. Genetic diversity evaluation of Cumin (*Cuminum Cuminum* L.) based on phenotypic characteristics. *Australian Journal of Corp science*, 5(3): 304-310.
- Baljeet, S.Y., Simmy, G., Ritika, Y., Roshanlal, Y. 2015. Antimicrobial activity of individual and combined extracts of selected spices against some pathogenic and food spoilage microorganisms. *International Food Research Journal*, 22(6): 2594-2600.
- Bialonska, D., Ramnani, P., Kasimsetty, S.G., Muntha, K.R., Gibson, G.R., Ferreira, D. 2010. The influence of pomegranate by-product and punicalagins on selected groups of human intestinal microbiota. *International J. Food Microbiology*, 140: 175-182.
- Dua, A., Gaurav, G., Balkar, S., Ritu, M. 2013. Antimicrobial properties of methanolic extract of cumin (*Cuminum cuminum*) seeds. *International Journal of Research in Ayurveda and Pharmacy*, 4(1): 104-107.
- Eikani, M.H., Golmohammad, F., Mirza, M., Rowshanzamir, S. 2007. Extraction of volatile oil from Cumin (*Cuminum cuminum* L.) with super-heated water. *Journal of Food Process Engineering*, 30: 255-266.
- Feldman, D.; Ganon, J.; Haffman, R., Simpson, J. 2003. *The solution for data analysis and presentation graphics*. 2nd Ed., Abacus Lancripts, Inc., Berkeley, USA.
- George, F.O., Ephraim, R.N., Lobasa, S.O., Bankole, M.O. 2009. Antimicrobial properties of some plant extracts on organisms associated with fish spoilage. University of Agriculture, PMB 2240, Abeokuta (UNAAB).
- Ibrahim, M.I., Husaini, A.S.A., Muhamad, N. 2016. Antimicrobial, total phenolic and total flavonoid properties of leaves and seed of *Jatropha curcas*, *Piper nigrum* L. and *Piper betle* methanolic crude extracts. *Malaysian Journal of Microbiology*, 12(6): 438-444.
- International Organization for Standardization "ISO" 1999. *International Organization for Standardization*. No. 6888-1:1999, A1:2003. *Microbiology of food and animal feeding stuffs — Horizontal method for the enumeration of coagulase-positive staphylococci (*Staphylococcus aureus* and other species) - Part 1: Technique using Baird-Parker agar medium (includes amendment A1:2003)*.
- International Organization for Standardization "ISO" 2001. *International Organization for Standardization*. No.16649-2. *Microbiology of food and animal feeding stuffs — Horizontal method for the enumeration of glucuronidase-positive *Escherichia coli* - Part 2: Colony-count technique at 44 °C using 5-bromo-4-chloro-3-indolyl-D-glucuronide*.
- International Organization for Standardization "ISO" 2017. *International Organization for Standardization*. No.6579-1. *Microbiology of the food chain Horizontal method for the detection, enumeration and serotyping of Salmonella - Part1: Detection of Salmonella spp.*
- Kaho, Z.M., Kadum, A.R., Hadi, A.A. 2019. Evaluation of antibacterial activity of *Piper nigrum* extract against *Streptococcus mutans* and *Escherichia coli*. *J. Pharmacological Science and Research*, 11(2): 367-370.
- Kaur, S., Noor, F., Swapanil, Y. 2017. Antibacterial activity of different extracts of black pepper. *International Research Journal of Advanced Engineering and Science*, 2(1): 172-173.
- Lukinmaa, S., Nakari, U., Eklund, M., Siitonen, A. 2004. Application of molecular genetic methods in diagnostics and epidemiology of borne bacterial pathogen. *APMIS*, 12, 908-929.
- Mostafa, A.A., Al-Askar, A.A., Almaary, K.S., Dawoud, T.M., Sholkamy, E.N., Bakri, M.M. 2018. Antimicrobial activity of some plant extracts against bacterial strains causing food poisoning diseases. *Saudi Journal of Biological Sciences*, 25: 361-366.
- Nagy, M., Socaci, S.A., Tofana, M. 2015. Determination of total phenolics, antioxidant capacity and antimicrobial activity of selected aromatic spices. *Bulletin UASVM Food Science and Technology*, 72(1): 82-85.
- Nasar-Abbas, S.M., Kadir, H.A. 2004. Antimicrobial effect of water extract of sumac (*Rhus coriaria* L.) on the growth of some food borne bacteria including pathogens. *International J. Food Microbiology*, 97: 63-69.
- Normanno, G., La Salandra, G., Dambrosio, A., Quaglia, N.C., Corrente, M., Parisi, A., Santagada, G., Firin, U.A., Crisetti, E., Celano, G.V. 2007. Occurrence, characterization and antimicrobial resistance of enterotoxigenic *Staphylococcus aureus* isolated from meat and dairy products". *Int. J. Food Microbiol.*, 115: 290-296.
- Pan, Y., Zhu Z., Huang Z., Wang H., Liang Y., Wang K., Lei Q., Liang M. 2008. Characterization and free radical scavenging activities of novel red pigment from *Osmanthus fragrans* seeds. *Food Chemistry*, 112: 909-913.
- Pandey, A. and Singh, P. 2011. Antibacterial activity of *Syzygium aromaticum* (Clove) with metal ion effect against food borne pathogens. *Asian J. Plant Sci. Res.* 1(2): 69-80.
- Rani, S.K., Saxena, N., Udaysree, S. 2013. Antimicrobial activity of black pepper (*Piper nigrum* L.). *Global Journal of Pharmacology*, 7(1): 87-90.
- Sapkota, R., Dasgupta, R., Nancy, Rawat, D.S., 2012. Antibacterial effects of plants extracts on human microbial pathogens & microbial limit tests. *International J. Pharmacological and Chemical Res.*, 2(4): 926-936.
- Shan, B., Cai, Y., Brooks, J.D., Corke, H. 2007. The in vitro antibacterial activity of dietary spice and medicinal herb extracts. *International J. Food Microbiology*, 117: 112-119.
- Sheikh, M.I., Islam, S., Rahman, A., Mostafizur, R., Mushiur, R., Muzanur, R., Abdur, A.F. 2010. Control of

- some human pathogenic bacteria by seed extracts of cumin (*Cuminum cyminum L.*). Agriculture Conspaction Science, 75(1): 39-44.
30. Singletary, K.W. 2010. Black Pepper: Overview of health benefits. Nutrition Today, 45(1): 43-47.
 31. Sodha, S.V., Griffin, P.M., Hughs, J.M. 2009. Food born disease. In: Mandell, G.L., Bennett, J.E., Dolin, R. (Eds). Principles and practice of Infectious Disease. 7th Ed. Philadelphia, Elsevier Churchill Livingstone: Chap. 99
 32. Sulieman, H.M.A., Allaaahmed, A.A.A. 2012. Effect of antimicrobial properties of pepper fruits on some spoilage organism of Sudanese wet-salted fermented fish (Fassiekh) product. World's Veterinary J., 2(1): 5-10.
 33. Zou, L., Hu, Y.Y., Chen, W.X. 2015. Antibacterial mechanism and activities of black pepper chloroform extract. J. Food Science and Technology, 52(12): 8196-8203.
- Staphylococci: implications for our food supply. Anim. Health Res. Rev., 13: 157-180.