

Microbiological evaluation of some heat treated fish products in Egyptian markets

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ABSTRACT

This study was conducted to confirm the bacterial conditions of fish products with E.O.S, and their hazards on public health. A total of 60 samples of fish products represented by smoked: herring and smoked salmon– semi cooked: fish finger and breaded shrimp (15 of each) were collected from different retail markets for bacteriological examination. The average of APC, *Coliform, Escherichia coli*, Mould & yeast and *Staphylococcus aureus* counts (log_{10} cfu/g) were 4.17± 0.12, 2.92±0.16, 2.19±0.23, 3.96±0.14 and 1.72±0.21 for herring, respectively, 3.16± 0.19, 2.69±0.13, 1.22±0.16, 2.22±0.18 and 1.06±0.06 in smoked salmon, 2.78±0.12, 2.02±0.22, 1.59±0.22, 2.14±0.15 and 1.24±0.24 in fish finger, respectively, and 2.60±0.13, 2.33±0.14, 1.46±0.23, 1.96± 0.20, 0± 0 in breaded shrimp, respectively. The incidence of some food poisoning bacteria (*Salmonella, Listeria monocytogenes* and *Vibrio parahaemolyticus* also investigated and no one of them was isolated in the examined samples.

Keywords: Salmon, Herring, Fish finger, Breaded shrimp

(http://www.bvmj.bu.edu.eg) (BVMJ-33(2): 305-316, DECEMBER, 2017

1. INTRODUCTION

Fishes are known to be highly nutritious and excellent sources of animal protein, which are consumed by larger percentage of the world's population because of its availability and palatability (Foran et al., 2005). Fish smoking is the most widely practiced and recommended method of preservation where sophisticated equipment for more improved methods is lacking. Smoking of food is achieved by lowering of the water activity via application of gentle heat. The surface of food which will normally support most commensal organisms is dried while the heat and chemicals inherent in the smoke deprives

microbes of necessary growth factors (Brown, 2004).

Prior to smoking, various pre-treatments, such as salting and drying, and/or after treatments, e.g. cooking and marinating, are applied in the industry. However, smoking is not an absolute preserving method. For this reason, the quality of raw material, the concentration of salt, water activity of the fish, heat through the smoking process, the quantity of smoke, the way of packaging, hygienic circumstances and heat of storage have important effects in reducing the risk of deterioration (Kaya and Erkoyuncu, 1999; Ahmed *et al.*, 2011).Today smoking is no longer "necessary", but it remains popular for the flavor it gives to such fish as salmon, tuna, trout etc.

Also, the processes of battering and breading provide special functions in food products including improving the appearance of the products, increasing the texture, reducing the oil uptake during the frying process and increasing the shelf life of the coated products (Varela and Fiszman, 2011). Battered and breaded fish products can undergo undesirable changes during frozen storage time due to microbial contamination from various sources and rapid spoilage as a result of protein denaturation (Benjackul et al., 2005) and lipid oxidation (Richard., 2002) leading to loss of quality. **Bacterial** contamination in food often results in food spoilage as well as life-threatening health hazards like food poisoning (Prescott et al., 1999).

Bacteriological examination is applied to evaluate the possible presence of microorganisms of public health significance and to give an impression about the hygienic quality of the fish. This includes temperature abuse and hygiene during handling and processing (Huss, 1995). Estimation of APC is used as an index in standards, guidelines and specifications and considered more useful to estimate spoilage and the remaining shelf life of fish and fishery products (Ólafsdóttiret et al., 1997).

In studies of seafood borne pathogens, four major pathogens have emerged as being of significant importance in terms of human health and disease. These include *Listeria monocytogenes, Vibrio parahaemolyticus, Staphylococcus aureus, and Salmonella spp.* (Feldhusen, 2000). L. monocytogenes has been isolated from fish and seafood products all over the world. V. parahaemolyticus is a human pathogen that occurs naturally in the marine environments and is frequently isolated from a variety of seafood including fish, shrimp, crab, lobster, scallop, and oyster (Austin, 2010). This pathogen is a common cause of foodborne illnesses in many Asian countries, including Taiwan, China, and Japan, and is recognized as the leading cause of human gastroenteritis associated with seafood consumption in the United States (Jaksic et al., 2002; Su and Liu, 2007).

These facts greatly influenced the interest of this study which aimed at assessing the microbial load of retailed smoked fish (herring and salmon) and some battered and breaded fish products (fish fingers and breaded shrimp).

2. Materials and methods

2.1. Collection of samples:

A total of 60 random samples of fish products (smoked: herring and smoked salmon - semi cooked: fish finger and breaded shrimp) (15 of each) were collected from different Giza supermarkets in Egypt. Each sample was kept in a separated sterile plastic bag and preserved in an ice box then transferred to the laboratory under complete aseptic conditions without undue delay and examined as quickly as possible. The collected samples were subjected to the bacteriological examinations to evaluate their safety and fitness for human consumption.

2.2. Preparation of samples (APHA (2001) :Ten grams from each sample were weighed and stomached with 90ml of 0.1% sterile buffered peptone water using stomacher (Seward stomacher 80 Biomasters, serial No 46464, England) to provide a dilution of 10-1. The homogenate was then allowed to stand for 15 minutes at room temperature. From the original suspension, one ml was transferred aseptically with sterile pipette into a test tube containing 9 ml of sterile buffered peptone water 0.1% and mixed well to produce a dilution of 10-2 from which further decimal serial dilutions were prepared. 2.3. Determination of Aerobic plate count (APHA, 2001)

2.4. Enumeration of Coliform bacteria & Escherichia coli (FDA, 2002)

2.5. Total Mould and Yeast Count (ISO 21527, 2008)

2.6. Isolation and Enumeration of Staphylococcus aureus (FDA, 2001)

2.7.Detection and Enumeration of Listeria monocytogenes (FDA,2011)

2.8. Isolation and identification of V. parahaemolyticus: According to (ICMSF, 1996)

3. RESULTS

It is evident from the result recorded in table (1) that APC in the examined samples varied from 3.54 to 4.97 with an average value of $4.17\pm0.12 \log \text{cfu/g}$, 1.00 to 3.92 with an average value of $3.16\pm0.19 \log \text{cfu/g}$, 2.04 to 3.72 with an average value of $2.78\pm0.12 \log \text{cfu/g}$ and 2.00 to 3.36 with an average $2.60\pm0.13 \log \text{cfu/g}$ for the examined samples of herring, salmon, fish finger and breaded shrimp, respectively. Table (2)

showed that the mean±SE of coliform and E.coli count of examined samples of herring, salmon, fish finger and breaded shrimp were 2.92±0.15 and 2.19±0.23, 2.69±0.13 and 1.22±0.16, 2.02±0.22 and 1.59±0.22 and 1.46 ± 0.23 , 2.33 ± 0.14 and respectively. Results achieved in table (3) indicated that the mean ±SE of moulds and yeast count of examined samples of herring, salmon, fish finger and breaded shrimp were 3.96±0.14, 2.22 ± 0.18 , 2.14 ± 0.15 and 1.96 ± 0.20 , respectively. It is evident from the results recorded in table (4) that the mean \pm SE of Staphylococcal aureus count of examined samples of herring, salmon, fish finger and breaded shrimp were 1.72±0.21, 1.06 ±0.06, 1.24 ± 0.24 and 0, respectively. Table (5) showed that the percentage and occurrence of Salmonella, Liesteria monocytogenes and Vibrio parahaemoliticus in examined samples of herring, salmon, fish finger and breaded shrimp based on their contamination were 0%, 0%, 0% and 0% of all investigated microorganisms respectively. Moreover, the results in table (6) showed that 100% and 100% of herring, salmon respectively, were unaccepted according to E.O.S (2005, 288). The results achieved in table (7) showed that 40%, 33%, 3.33% of fish finger and breaded respectively shrimp were unaccepted according E.O.S (2005,3495). to

	Smoked fish		Semi- cooked		
No. of positive samples	Herring	Salmon	Fish finger	Breaded shrimp	
	15	15	15	15	
%	100	100	100	100	
Mini.	3.54	1.00	2.04	2.00	
Maxi.	4.97	3.92	3.72	3.36	
Mean	4.17	3.16	2.78	2.60	
SE	0.12	0.19	0.12	0.13	

Table (1): Statistical analytical results of Total aerobic plate count log cfu/g in fish products samples

Table (2): Statistical analytical results of Coliform and E. coli counts log cfu/g in fish products samples

No. of	Sı	moked fis	h		Semi-	cooked		
po siti ve	Herring		Salmon		Fish finger	Bre	aded shrimp	
sa mp	Coliforms	E. coli	Coliforms	E. coli	Coliforms	E. coli	Coliforms	E. coli
les	14	4	6	6	11	5	5	5
%	93	27	40	40	73	33	33	33
Min.	2.04	1.70	2.18	1.00	0.04	1.00	2.04	1.00
Max.	3.80	2.60	2.97	2.00	2.79	2.18	2.79	2.00
Mean	2.92	2.19	2.69	1.22	2.02	1.59	2.33	1.46
SE	0.16	0.23	0.13	0.16	0.22	0.22	0.14	0.23

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Smoke	Smoked fish		oked
Herring	Salmon	Fish finger	Breaded shrimp
15	10	11	10
100	67	73	67
3.11	1.40	0.79	1.15
4.72	2.97	2.63	3.11
3.96	2.22	2.14	1.96
0.14	0.18	0.15	0.20
	Herring 15 100 3.11 4.72 3.96	HerringSalmon1510100673.111.404.722.973.962.22	HerringSalmonFish finger15101110067733.111.400.794.722.972.633.962.222.14

Table (3): Statistical analytical results of Mould and yeast count log cfu/g in fish products samples (N = 15)

Table (4): Statistical analytical results of Staphylococcus aureus count log cfu/g in fish products samples (N = 15 each)

No. of positive	Smoked fi	sh	Semi- cooked		
samples	Herring	Salmon	Fish finger	Breaded shrimp	
	12	5	2	0	
%	80	33	13	0	
Mini.	0.70	1.00	1.00	-	
Maxi.	2.78	1.30	1.48	-	
Mean	1.72	1.06	1.24	-	
SE	0.21	0.06	0.24	-	

Table (5): Frequency and	percentage occurrence	of bacterial isolates	of fish products san	ples

	Smoked fish			Semi-cooked		
Isolates	Herring	Salmon	Fish	finger	Br. shrimp	
	No %	No %	No	%	No %	
Salmonella spp.			-	-		
Listeria monocytogenes				-		
Vibrio parahaemolyticus			-	-		

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	Acceptable limits	Herring	Salmon
		Non Accepted	Non Accepted
		N/15 %	No/15 %
APC	$\leq 10^5$	0 0	0 0
Coliforms	≤ 10	14 93	6 40
Moulds	Free	15 100	15 100
E.coli	Free	4 27	6 40
Listeria, monocytogenes	Free	0 0	0 0
Salmonella	Free	0 0	0 0
Staph aureus	Free	11 73	5 33
Vibrio parahaemoleticus	Free	0 0	0 0

Table (6) Acceptability of the examined samples of smoked fish samples according to EOSQC (2005/288)

Table (7) Acceptability of the examined samples of smoked fish samples according to EOSQC (2005/3495)

	Acceptable limits	Fish finger Non Accepted		Bread	Breaded shrimp Non Accepted		
				Non			
		N/15	%	N/1	5 %		
APC	$\leq 10^5$	0	0	0	0		
Coliforms	$\leq 10^2$	9	60	5	33		
Moulds	Free	11	73	15	100		
E.coli	Free	5	33	5	33		
Listeria, monocytogenes	Free	0	0	0	0		
Salmonella	Free	0	0	0	0		
Staph aureus	Free	2	13	0	0		
Vibrio parahaemoleticus	Free	0	0	0	0		

4. DISCUSSION

Fish is a reservoir of large number of microorganisms; one of the major factors contributing to poor quality of the fish in retail trade is unhygienic handling and storage leading to off smell, physical damage and contamination with dirt and objectionable microorganisms (Sugumar et al., 2004). Eyo, 2001 stated that microbial action has been known to play a large part in the spoilage of fish. Bacterial spoilage is characterized by softening of the muscle tissue and the production of slime and offensive odors.

1- The aerobic plate count

The aerobic plate count (APC) is an important factor for evaluation of microbial quality assessment in food products and is an indicator of the overall degree of microbial contamination of foods (ICMSF, 1986). APC does not measure the entire bacterial population but rather the number bacteria growth in the presence of oxygen (aerobically) medium and at range (mesophilic) temperatures.

Table (1) revealed that, the mean value of aerobic plate counts APC (\log_{10} cfu/g) mean±SE in the examined smoked fish (smoked fish (Herring - Salmon) and semicooked fish products (Fish finger --Breaded shrimp) were (4.17 ±0.12--3.16± 0.19) and $(2.78 \pm 0.12 - 2.60 \pm 0.13)$ respectively. Higher findings were observed by Khater and Farag (2016) who found the APC in herring and salmon paste samples were 5.35 ± 0.23 and 5.34 ± 0.68 respectively, also Ibrahim et al., 2014) found the APC in was 2.06×10^6 cfu/g. The smoked fish bacterial load were found to be higher in the smoked fish samples which might be due to secondary contamination during the time of handling as well as storage of fishes in ice

made from contaminated water, poor hygiene and sanitation condition of processing (Hatha et al. 1998). Smoking helps in inhibiting the activities of microorganisms, however, when smoking process not properly carried out, microbial growth and activities still continue, leading to the deterioration of the fish. Thus, TAC is considered a quality indicator for food. Although there is not direct correlation between this and the presence of pathogenic microorganisms, TAC is an indicator of the shelf-life of products, and also the potential for growth of the microorganism that is present (Arvanitoyannis et al., 2005). Our lower findings observed in fish finger and breaded shrimp may be due to adding garlic and pepper powder and other spices caused to reduce the bacterial count in fish fingers due to their antibacterial role. Higher results were seen in Ibrahim-Hemmat et al, (2015) who found the APC of fish finger was $8.33 \times 10^4 \pm 1.04 \times 10^4$ cfu /g.

2- Coliform and E.coli count

The results recorded in table (2) revealed that, the mean value of Coliform and Escherichia coli count in (smoked fish (Herring --Salmon) and semi cooked fish products (Fish finger -- Breaded shrimp) were $2.92\pm$ $0.16, 2.19 \pm 0.23$ for herring, 2.69 ± 0.13 , 1.22±0.16 for salmon and 2.02 ± 0.22 , 1.59 \pm 0.22 for fish finger and 2.33 \pm 0.14, 1.46 ± 0.23 for breaded shrimp. The Coliform counts were low in semi-cooked fish products; this may be due to the attained temperature for frying was sufficient to kill vegetative bacteria.

Data presented in Table (2) showed that, the mean value of coliform count came in parallel with those of (Soliman *et al.*, 2002,

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Abd El-Rahman *et al.*, 2003, Vigano *et al.*, 2007). Munce (1980) stated that presence of *Coliform* in food has been linked with the practice of inadequate hygienic measure, mishandling, improper storage and use of dirty water during marketing and all unhygienic condition of the shops.

3-mould and yeast count

The incidence of mould in fish could be attributed to improper sanitation during catching, handling, processing, salting, storage, transportation, distribution and marketing of fish. Contamination with a variety of mould species resulted in undesirable changes of fish and rendering it unfit for marketing and increase the risk of infection with respective disease to consumers as a probable result of aflatoxins production by some fungal strains. The results recorded in table (3) revealed that, the mean value of mould and yeast counts (log10 cfu/g) mean±SE in the examined smoked fish (Herring -- Salmon) and semicooked fish products (Fish finger -- Breaded shrimp) were 3.96 ± 0.14 , 2.22 ± 0.18 , 2.14 ± 0.21 and $1.96 \pm$ 0.20, respectively. Cold-smoked fish are not cooked, because the temperature generally does not exceed 43°C. Therefore, the most common causes of spoilage in smoked fish are mold growth .The count of molds and yeast.

In Herring and Salmon respectively. Similar results have been reported by Tadros - Safaa (1999) who found that the mean value of total mould count was 7.5 $X10^2 \pm 2.4 X10^2$ /g of smoked fish. Nearly similar findings obtained by Ibrahim –Hala (2000) who reported that the mean value of the total mould count/g of smoked fish was $3.5X10^3 \pm 1.3 X10^3$. Also (El-Sayed, 1995) reported that the mean value of total mould count /g of smoked fish was $15.3X10^3$.

4- Staphylococcus aureus count

Staphylococcus aureus is a major cause of poisoning due to ingestion food of enerotoxins (Stengel, 1990); the ability to produce such enterotoxins in food is more likely when competing microorganisms were absent (Frazier and Westhoff, 1984). It is evident from the results recorded in table (4) revealed that the incidence of staphylococcus aureus were 80, 33, 13 and 0 % in the examined smoked fish (Herring -- Salmon) semi-cooked (fish finger and breaded shrimp) with an average 1.72±0.21and 1.06±0.06 in herring and smoked salmon, respectively. While in examined semi-cooked fish products fish finger was 1.24±0.24 and breaded shrimp samples were free from coagulase positive Staph. aureus. Also our results were in agreement with Adeyeye et al, (2016) who found the mean S. aureus count of smoked fish 1.1 ± 10^2 to 3.8 ± 10^2 cfu/g whereas Zaki (2003) recorded 3 log cfu/g Staphylococcus aureus count in smoked fish which was higher than our results. The presence of Staphylococcus aureus in smoked samples be attributed to post-processing can Contamination. Our results were agreed with those of (Ahmed and Anwar, 2007 and Abd Allah, 2010) who found that all examined shrimp samples were free from coagulase positive S. aureus.

5-Prevalence of food poisoning organisms (Salmonella, Listeria monocytogenes and Vibrio parahaemolyticus

The results recorded in table (6) revealed that none of the three food poisoning organisms were detected in the examined fish products samples.

a) Salmonella

Salmonella was not detected in the samples analyzed in this study, which was in

agreement with previous studies (Sulieman et al. 2014) in seafood products. Meanwhile, disagreed with those of Soliman *et al.*, (2002), Younis, (2013) who isolated *Salmonella* from fried fish, shrimp.

b) Vibrio-parahaemolyticus

V.parahaemolyticus is an indigenous bacterium in the marine environment and can also grow in 1-8% salt (Khodaeeyan, 2008, Akhonzade Basti et al., 2006). *Salmonella* spp. and *V.* parahaemolyticus in aquaculture products mainly originates of hygiene and sanitation. But sometimes, incidence of these bacteria in fish may occur due to external contamination. Fortunately no presence of pathogenic *Vibrio-parahaemolyticus* were found in all inspected fish products samples

c) Liesteria monocytogens

In the present study, *L. monocytogenes* not detected in all examined samples. Similar results observed in Jalali and Abedi (2008) they don't found *L. monocytogenes* in 85 samples of fresh and frozen fish and shrimp analyzed. *L. monocytogenes* contamination of seafood varies with product category. Jorgensen and Huss (1998) demonstrated that

5. CONCLUSION

Finally, the study concluded that smoked fish, which are ready for immediate human consumption, have unacceptable microbial quality. However, they may consider of highrisk due to fungal toxins hazards.

6. REFERENCES

Abd El-Rahman, A.A., Hamed, N.A., El-Timawy, A.M. and Kaldes, Y.T. 2003. Bacteriological evaluation of some foodborne pathogenic bacteria the highest prevalence of *L.monocytogenes* is in cold-smoked fish (34%-60%), whereas the lowest is in heat-treated and cured seafood (4%-12%). In general, L. monocytogenes is not usually found on fish captured from open waters. However, contamination may take place long before the fish raw material reaches retail trade or processing factories. Potential sources of L. monocytogenes on fishing vessels include contamination from water and ice, soiled surfaces, and boxes as well as from human and avian sources. As L. monocytogenes is commonly found in coastal waters and in surface waters of lakes, fish captured or cultivated in these waters may possibly carry this microorganism (FAO, 1999).

Table (6) showed that 100%, 100% were unaccepted based on their moulds &yeasts count/g according to E.O.S (2005) of examined samples of herring and smoked salmon respectively. Results achieved in table (7) indicated that 73% and 67% of the examined fish finger and breaded shrimp samples were unaccepted based on their moulds & yeasts count/g according to E.O.S. (2005)

So, special attention should be taken from competent authorities and food business operators. Moreover, consumers are increasingly aware of the danger of pathogens in RTE fish. Also, the present study proved that semi-cocked are considered of public health hazard due to the presence of considerable percentages of *coliform*.

transmitted by grilled, fried fish.Egypt. J. Agric. Res., 811:383-396.

Abd Allah, M.S. 2010. Microbiological risk assessment in raw, ready-to-eat fish

at Dakahlia province. Ph.D.Thesis, Food Hygiene &Control, Fac. Vet. Med., Mansoura Univ.

- Adeyeye, S.A.O., Oyewole, O.B., Obadina,
 A.O., Omemu, A.M., Adeniran, O.E.
 and Oyedele, H.A. 2016. Assessment
 of quality and safety of traditional
 smoked spotted tilapia fish (Tilapia
 mariae) from Lagos State, Nigeria.
 Nutrition & Food Science 46, 142155.
- Ahmed, S. and Anwar, M.N. 2007. Bacteriological assessment of value added ready to cook/ eat shrimps processed for export from Bangladesh following the guidelines of international standards. Bangladesh J. Microbiol., 242: 81-84.
- Ahmed, A., Dodo, A., Bouba, A.M., Clement,
 S. and Dzudie, T. 201. Influence of traditional drying and smoke-drying on the quality of three fish species (Tilapia nilotica, Silurus glanis and Arius parkii) from Lagdo Lake, Cameroon. J. of Animal and Veterinary Advances, 10 (3)301-306.
- Akhondzadeh, A., Misaghi, A. and Kamkar, A. 2006. Bacterial pathogens in fresh, smoked and salted Iranian fish. Food Control. 17 (3) 183-188.
- American Public Health Association (APHA) 2001. Compendium of methods fo the microbiological examination Of Food. 4th ed., Washington, D.C.
- Arvanitoyannis, I.S., Tsitsika, E.V. and Panagiotaki, P. 2005. Implementation of quality control methods(physicochemical,
 - microbiological and sensory) in conjunction with multivariate analysis towards fish authenticity. International

Journal of Food Science and Technology 40, 237-263.

- Austin B. 2010. Vibrios as causal agents of zoonoses. Vet Microbiol. 140:310–317.
- Benjakul, S., Visessanguan, W., Thongkaew, C. and Tanaka, M. 2005. Effect of frozen storage on chemical and gelforming properties of fish commonly used for surimi production in Thailand. Food Hydrcolloids, 10: 197-207.
- Brown G. E. 2004. A Report on the Prevalence of Bacteria specie in Retailed Smoked Fish within Bauchi Metropolis. El-Sayed, Y. S. A. 1995. mycological studies on Locally produced smoked fish with special reference to toxigenic strains.Ph.D. Thesis, Dept.Food Control. Fac. Vet. Med. Zagazig University.
- EOSQC (Egyptian Organization for Standardization and Quality Control), 2005. 889-2/2005 for fish and fish product.
- Eyo A. A. 2001. Fish Processing Technology in the Tropics. 1- 20 pp. Mendez E, Gonzalez RM, Inocente G, Giudice H, Grompone MA (1996) Lipid content and fatty acid composition of fillets of six fishes from the Rio de la Plata. J. Food Compos. 9(2): 163-170.
- [FAO] Food and Agriculture Organization. (1999)Fisheries Report No. 604. Expert Consultation on the Trade Impact of Listeria in Fish Products. Amherst, MA.
- Feldhusen F. 2000. The role of seafood in bacterial food-borne diseases. Microbes Infect .2:1651–1660.
- Food and Drug Administration (FDA) 2001. U. S. Department of health and human

services. Public Health Service. Food and Drug Administration. College Park, MD 20740.

- Food and Drug Administration (FDA) 2002. Enumeration of *Escherichia coli* and the coliform bacteria: Bacteriological Analytical manual. Chapter 4.
- Food and Drug Administration (FDA) 2011. Quantitative Assessment of Relative Risk to Public Health from Foodborne Listeria monocytogenes Among Selected Categories of Ready-to-Eat Foods. Fish and Fishery Products Hazards and Controls Guidance Fourth Edition . appendix 5 : FDA and EPA Safety Levels in Regulations and Guidance.
- Foran, J.A., carpenter, D.D., Hamlton, M.C., Knuth, B.A. and Schwager, S.J. 2005. Risk based consumption advice for farmed Atlantic and wild pacific salmon contaminated with dioxin and dioxin-like compound. Environmental Health Perspective. 33: 350-356.
- Frazier, W.C. and Westhoff, D.C. 1984.Tata McGraw Hill publisaing Co. Limited New York .U.S. A.
- Mohamed, A. A., Hatha, N. Paul and Rao, B. 1998. Bacteriological quality of individually quick-frozen (IQF) raw and cooked ready to-eat shrimp produced from farm raised black tiger shrimp (Penaeus monodon) Food Microbiology. 15, 177–183.
- Hemmat M. Ibrahim, Reham A. Amin, NahlaA. Shawky , Suzan, H. Sheir 2015.Aerobic spore formers in battered and breaded fish products. Benha VetrinaryMedical J. 28 (2):123 128.

- Jaksic, S., Uhitil, S., Petrak, T., Bazulic D., and Karolyi LG. 2002. Occurrence of Vibrio spp. in sea fish, shrimp and bivalve mollusks harvested from Adriatic Sea. Food Cont.13:491–493.
- Jalali, M. and Abedi, D. 2008. Prevalence of *Listeria* species in food products in Isfahan, Iran. Int J. Food Microbiol. 122:336–340.
- Jorgensen ,L.V. and Huss H.H. 1998. Prevalence and growth of *Listeria monocytogenes* in naturally contaminated seafood. Int J Food Microbiol. 42:127–131.
- Huss, H.H. 1995. Quality and quality changes in fresh fish. FAO Fisheries Technical Paper 348 FAO. Rome, Italy.
- Ibrahim A. M. Hala 2000. Incidence of fungal contaminants in fish and fish products.M.V.Sc. Thesis, Dept .Meat Hygiene, Fac. Vet. Med . Zagazig Univ . Benha branch.
- International Commission on Microbiological Specification for Foods "ICMSF": 1996. Vibrio parahaemolyticus. Microorganisms Foods. in Characteristics of Microbial Pathogens Blackie Academic & Professional. 426-435. London. International Committee Microbiological on Specification for foods.
- ICMSF, 1996. Micro-organisms in foods 5. characteristics of Microbial Pathogen, Blachie Academic &Professional, London (ISBN0412 47350 X).
- ISO 21527-1. 2008. Microbiology of food and animal feeding stuffs - horizontal method for the enumeration of yeasts and moulds -- part 1: Colony count technique in products with water activity greater than 0.95.

- Kaya,Y. and Erkoyuncu, I. 1999. Deg`is,ik
 Dumanlama Metotlarının Balık
 Turlerinin Kaliteleri Uzerine Etkisi. O.
 M.U, Ziraat Fakültesi Dergisi, Vol. 14
 No. 1, 93-105.
- Khater, F. Dalia and El-Safy, F. Samia 2016. Evaluation of bacterial and chemical quality of new manufactured pasted fish products in a large scale fish processing plant, Egypt. Benha Veterinary Medical J. 31 (2):63-72.
- Khodaeeyan Ghegeni, F., 2008. Classification of bacteria. Food Microbiology. 9-11, 138-140.
- Munce, H. R. 1980. Principles of food packaging an international _ guide/FAO, United Nation. pp. 19-210lafsdóttir, G., Martinsdóttir, E., Oehlenschläger, J., Dalgaard, Р., Jensen, B., Undeland, I., Nilsen, H. 1997. Methods to evaluate fish freshness in research and industry.Trends in Food Science &Technology, 8:258-265.
- Prescott, L.M., Harry, J.P. and Klein, D.A. 1999. Food and Industrial Microbiology. Chapter 43, 4thEdition, Newyork, McGraw Hill publication.
- Richards, M.P. 2002. Contributions of blood and blood components to lipid oxidation in fish muscle. PhD Thesis, University of Massachusetts, Amherst, USA.
- Soliman, M.R., Abd El-Monem, K.H. M. and Saad, S.M. 2002. Microbiological quality of ready to eat meat product, fishes in urban, rural qrean. J. Egypt. Vet. Med. Ass., 626:39:51.

- Stengel, G.F. 1990. *Staphylocooci*, Fleisch wirtschaft 70 (3): 307-312.
- Sulieman, A.E., Hassan, Z.M.A., Elkhalifa, E.A., 2014. Microbial Safety of Dried Fish Meat Kejeik Produced in Sudan. Food and Nutrition Sciences 5, 606-613.
- YC Liu C. Su, and 2007. Vibrio parahaemolyticus: concern a of seafood safety. Food Microbiol. 24:549-558.
- Tadros Safaa, S. 1999. Mycological contamination of some Fish products at Alexandria province. M.V.Sci. Thesis, Dept. Food Control. Fac. Vet. Med. Alexandria Univ, Egypt.
- Varela, P., Fiszman, S.M. 2011. Hydrocolloids in 36:647-655. fried foods. A review. Journal of Food Hydrocolloids, 25:1801-1812.
- Vigano, A., Pellissier, N., Hamad, H.J., Ame, S.A. and Pontello, M. 2007: Prevalence of E. coli, Thermotolerant *Coliforms*, *Salmonella spp.Vibrio spp*. in ready-toeat foods: Pemba Isl, United Republic of Tanzania.Ann.lg., 195:395-403.
- Younis, A. Enas, 2013. Studies on incidence of *Salmonella spp., Listeria monocytogenes* to gives in some of ready to eat foods. M.V.SC. Thesis Microbiology, Fac. Vet .Med. Cairo Univ.
- Zaki, M. Eman, 2003. Risk assessment of ready prepared meat products. Ph. D. Thesis, Meat Hygiene, Fac. Vet. Med., Cairo University.