



## Determination of the optimal protective dose of inactivated *Salmonella Typhimurium* vaccine in pigeon

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### ABSTRACT

Salmonellosis is one of the major health problem affect pigeons and protection of pigeons against this disease is very important, for this purpose the present work was planned to prepare an inactivated Montanide ISA- 206 oil adjuvant vaccine from a locally pigeon isolate of *Salmonella Typhimurium* (*S. Typhimurium*) and determined of the optimal dose, which protect pigeon against salmonellosis. Evaluation of such preparation following the quality control tests revealed that it was free from any foreign contaminants, safe and immunogenic. Pigeons were divided into six groups, three of them vaccinated subcutaneously with single dose (0.5ml) of different bacteria concentrations ( $10^8$ ,  $10^9$  and  $10^{10}$  CFU/dose) and the others three received 2 doses with 3 weeks intervals with same route and concentrations. The vaccination/challenge assay with a virulent *S. typhimurium* organism using  $5 \times 10^7$  CFU/dose revealed that protection rates of vaccinated birds were 70% ,75% and 75% for the first three groups (Ia, IIa, IIIa) respectively, while the other three groups gave 80%, 85% and 85% protection (Ib, IIb, IIIb) respectively. Control groups could not withstand the challenge and protection rate were 15% and 20% respectively. The seroevaluation showed that the humoral immune response developed against *S. typhimurium* in vaccinated pigeons was high. On conclusion the optimal dose of the prepared vaccine should be  $10^9$  CFU/dose applied twice with three weeks interval, which can cover the needed requirements and protect pigeon against salmonellosis.

**Keywords:** *S. Typhimurium*, pigeon, vaccine, humoral immune response

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### 1. INTRODUCTION

*Salmonella* are host-adapted, zoonotic human and animal pathogens (Libby et al. 2004). The disease caused by this organeasm in pigeons is a major bacterial disease, resulting in mortality in squabs and occasional deaths in adult in infected pigeon lofts. More or less severe clinical signs, including weight loss, diarrhoea, polyuria, lameness and inability to fly are more common. Treatment of infected flocks is difficult since even long-term antibiotic therapy may leave subclinical carriers that keep the *Salmonella* infection in the loft going (Monita Vereecken et al., 2000). Bacterin vaccines proved unable to prevent initiation of *Salmonella* infections in pigeons (Uyttebroek et al., 1991) and vaccinated birds developed clinical signs and lesions during infection. For this reason, effective prevention of salmonellosis in pigeons through vaccination is not yet available. Recently, a newly developed *Salmonella* bacterin vaccine has been marketed. Duchatel et al. 1998 demonstrated a significant decrease of mortality in pigeons vaccinated with this vaccine following intramuscular challenge

with *Salmonella typhimurium* var. Copenhagen. The objectives of the present study were to prepare an inactivated oil-emulsion bacterin from a local pigeon isolate of *Salmonella typhimurium* and evaluation of this vaccine through determination of the optimal dose ,evaluate its efficacy for protecting pigeon against salmonellosis under experimental conditions and monitoring the humoral immune response in vaccinated and unvaccinated birds..

### 2. MATERIALS AND METHODS

#### 2.1. Pigeons:

A total of two hundred and seventy 4 weeks age Pigeons obtained from a commercial pigeon-breeding centre, Kept under strict hygienic measure of rearing and feeding. Cloacal swabs and blood samples were collected from pigeon to confirm that they were free from *Salmonella typhimurium* bacteria or antibodies.

#### 2.2. Vaccine preparation

A local pigeon isolate of *Salmonella typhimurium*, was kindly obtained from Central laboratory for evaluation of veterinary biologics (CLEVB). This strain was identified morphologically, culturally, biochemically and serologically following the method adapted by (Nagraja et al. 1991). The strain was grown on tryptic soya agar (Difco) for 48 hr at 37° C. Bacteria was harvested in normal saline, and the concentration of bacterial suspension was adjusted to contain 10<sup>8</sup>, 10<sup>9</sup> and 10<sup>10</sup> CFU/ dose. The purity of culture was examined by inoculation on to brilliant green agar plate (Difco). The bacterial culture was inactivated by adding 0.3% formalin

with agitation then the inactivation was ensured by plating onto nutrient agar and incubation at 37°C for 24 hrs. Montanide ISA- 206 was added to the different concentration cultures in equal volume (v/v) and mixed thoroughly according to (Steward, 1983).

### 2.3. Experimental Design:

Two hundred and seventy pigeons were used in this experiment. 30 pigeons were used for the safety testing while the rest (240 pigeons) were divided into different groups as following table.

Table (1): Experimental Design:

GROUPS	No of pigeon	Bacteria concentration	1st dose	Booster dose	Challenge
Ia	30	10 <sup>8</sup>	Each bird in each group received 0.5 ml S/C from each concentration	-	20 pigeons from vaccinated and control groups challenged with 5 x 10 <sup>7</sup> CFU/dose three weeks post 1st dose or boosting, bird observed for 14 days for any clinical symptoms or mortality
Ia	30	10 <sup>9</sup>		-	
IIIa	30	10 <sup>10</sup>		-	
Iva	30	Normal saline		-	
Ib	30	10 <sup>8</sup>		-	
IIb	30	10 <sup>9</sup>		-	
IIIb	30	10 <sup>10</sup>		-	
Ivb	30	Normal saline		-	

N.B: Ten pigeons from each concentration in each group were left without challenge for seroevaluation (group Ia, IIc and IIIc) for 3 weeks in group vaccinated with single dose and for 7 weeks in groups vaccinated with booster dose.

### 2.4. Evaluation of locally prepared inactivated Salmonella typhimurium vaccines

#### a. Sterility tests:

The prepared vaccines were tested for freedom from any bacteria, fungi and mycoplasma contaminant, according to (OIE, 2012).

#### b. Safety test:

Ten birds were inoculated with the double field dose s/c of each concentration (10<sup>8</sup>, 10<sup>9</sup> and 10<sup>10</sup> CFU/ dose) and kept under daily observation for 14 days according to (OIE, 2012)

#### c. Potency tests:

Protection rates of the prepared Montanide ISA 206 vaccine of *Salmonella typhimurium* vaccine containing 10<sup>8</sup>, 10<sup>9</sup> and 10<sup>10</sup> CFU/ dose were carried out primarily by using the vaccination/challenge test.

#### 1. Serological evaluation:

Seroevaluation of immune response was adopted for all groups where serum samples were collected from vaccinated as well as control group and humoral immune response developed against *S. Typhimurium* tested by microagglutination test according to (Brown et al. 1981).

#### 2. Challenge test:

Twenty pigeons from vaccinated and control groups were evaluated through vaccination/challenge test by inoculation 0.1 ml intramuscularly of virulent strain of *Salmonella typhimurium* serovar typhi in a concentration of  $5 \times 10^7$  CFU/dose three weeks post first dose or booster dose of the prepared vaccine as in table(3). The virulent strain was obtained from CLEVB, Abassia, Cairo, Egypt . Birds were kept under observation three weeks post challenge exposing mortality and disease symptoms OIE Manual (2012).

2.5. Reisolation of challenge strain from challenged birds:

Samples were taken from internal organs (liver, heart and spleen) of mortality cases and birds showed symptoms of salmonellosis post-challenge. These samples were preenriched then

streaked onto MaConkey and *Salmonella shigella* agar media.

3. RESULT

Regarding the humoral antibody responses, it was checked using micro-agglutination test and the antibody titers were calculated as shown in Table (2). The level of antibody in sera of pigeons vaccinated with local inactivated vaccines of *Salmonella typhimurium* increased in group I, II and III, 3 weeks post the 1st dose of vaccination and reach to maximum at the 5<sup>th</sup> week post boosting in the same groups. The protection efficacy of the local prepared *Salmonella typhimurium* vaccines in pigeons are shown in table (3).

Table (2): Mean antibody titer of microagglutination test in the pigeons vaccinated with locally prepared inactivated montanied ISA 206 *Salmonella typhimurium* vaccines.

Sub-groups	Pre-vaccinatio	Weeks post 1st dose			Weeks post boosting						
		1	2	3	1	2	3	4	5	6	7
(Ic)	0	4	8	32	32	64	128	128	128	128	64
(IIc)	0	8	32	64	64	128	128	128	256	128	128
(IIIc)	0	16	32	64	64	128	128	256	256	128	128
Control unvaccinated	0	0	0	0	0	0	0	0	0	0	0

Table (3): Protective percentage of local mantonide ISA 206 *Salmonella typhimurium* vaccines in pigeon challenged with virulent *S. typhimurium* strain.

Groups	Protection Percent post challenge								
	Mortality	1st dose Survival		Protection %	Sub-groups	Mortality	Boostering Survival		Protection %
		With lesions	With out lesions				With lesions	Without lesions	
(Ia) (10 <sup>8</sup> /dose)	3/20	3/20	14/20	70 %	(Ib) 10 <sup>8</sup> /dose	2/20	2/20	16/20	80 %
(IIa) (10 <sup>9</sup> /dose)	3/20	2/20	15/20	75%	(IIb ) 10 <sup>9</sup> /dose	0/20	3/20	17/20	85 %
(IIIa) (10 <sup>10</sup> /dose)	4/20	1/20	15/20	75%	(IIIb) 10 <sup>10</sup> /dose	1/20	2/20	17/20	85 %
Control unvaccinated	7/20	10/20	3/20	15%	control	6/20	10/20	4/20	20 %

#### 4. DISCUSSION

It is widely known that the number of cases of Salmonella related food poisoning have increased year after year over the past decade. Salmonellae are responsible for considerable losses in the poultry industry through the death of birds and loss in production and it is estimated to cost poultry farmers in some countries (Hassan et al., 2013). Vaccination is one of the most important methods for prevention of Salmonellosis. There has been an increasing interest in using Salmonella vaccination in poultry especially against the serovars of major public health relevance; *S. Enteritidis* and *S. Typhimurium*. Inactivated and/or live Salmonella vaccines are in use for poultry in a number of countries (Vielitz et al., 1992). Many racing pigeons carry Salmonella asymptotically in their bowels. The stressors associated with racing can act as trigger factors enabling the bacteria to penetrate the bowel wall and spread throughout the body causing a range of symptoms that compromise race performance. For control of Salmonella infected pigeon flocks, not only is the clinical disease to be avoided, but it is also necessary to inhibit excretion of the organism since this could maintain the infection (Monita et al., 2000). The locally prepared *S. Typhimurium* vaccine in this study was free from bacterial, fungal and mycoplasma contaminant and it was safe when injected in a doubled dose in pigeons for all concentration used.

Regarding the humoral antibody responses, it was checked using micro-agglutination test and the antibody titers were calculated.

The level of antibody in sera of pigeons vaccinated with local inactivated vaccines of Salmonella typhimurium increased from 0 titer pre-vaccination to be 32, 64, and 64 against Salmonella typhimurium in the group Ic, Iic and IIIc respectively 3 weeks post the 1st dose of vaccination and reach to maximum at the 5<sup>th</sup> week post boosting in the same groups (128, 256 and 256) respectively. These results coincide with that proved by Nagraja et al. (1991).

Challenge test is considered the master test for determination of the protective value of a vaccine (Timms et al., 1990). Concerning the protection efficacy of the local prepared *Salmonella* typhimurium vaccines in pigeons, the data showed that protection rates for pigeons received single dose were 70 % 75 % and 75% in the group Ia, IIa and IIIa respectively. While the protection rate was 80 % 85 % and 85% in the group Ib, IIb and IIIb respectively for pigeons received doubled dose.

While control group IV gave only 15 % and 20% protection under the same conditions.

The achieved protection values by the prepared vaccines are accepted to pass the vaccine for use according to Egyptian Veterinary Codex- CLEVB (2009).

The challenge strain was re-isolated from internal organs of all mortality cases and pigeons showed symptoms of salmonellosis post-challenge.

The prepared inactivated formalized *S. typhimurium* vaccine was safe and effective and that agree with (Barrow 1991) who stated that inactivated vaccine of avian salmonellosis is protective against Salmonella infection. Also these results came confirming by those of Timms et al. (1990) and Uyttebroek et al. (1991) who found that formalized *S. typhimurium* vaccine protected pigeons against experimental challenge. From above obtained data in this study it could be concluded that the optimal dose of the prepared vaccine should be 10<sup>9</sup> CFU/dose applied twice with three weeks interval, which can cover the needed requirements and protect pigeon against salmonellosis.

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