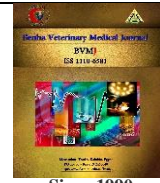




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Review Article

Effects of prebiotics supplementation on Nile tilapia, emphasizing phospholipids aquafeed - A review.

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ABSTRACT

Aquaculture has been a growing provider of animal protein industry in recent decades. Tilapia is a very valued fish cultivated across a vast geographical region in numerous countries, including Egypt. Tilapia has the advantage of being tolerant of a wide range of environmental circumstances and utilizing food from the lowest trophic levels. In recent decades, aquaculture has become a greater contribution to animal protein supply. Tilapia is a highly valued fish grown in a wide range of nations, including Egypt, as a source of animal protein. Tilapia could be tolerant to various environmental conditions. Some natural bio-friendly feed additives such as probiotics, prebiotics, and synbiotics are become a new approach in aquaculture industry to maintain aquaculture species health and a good environment at the same time produce chemical and antibiotic-free aquaculture product they become popular dietary supplements with the potential to increase not only growth performance, but also immunological competence and general well-being of fish and crustaceans. The prebiotics preparations are known to be important feed additives to be used in the aquaculture industry to sustain both aquatic species and environmental health. This makes them popular incorporations to enhance not only the growth indicators but also the immunological competence and general well-being of fish and crustaceans. The current review paper focuses on the possible effects of phospholipids (P.L) containing prebiotics on the health, immunological response, growth performance, and survival of *oreochromis niloticus*.

1. *Oreochromis niloticus* culture

The Nile tilapia *Oreochromis niloticus* is typically found throughout the African countries and worldwide (Elsaid et al., 2019). It is a member of cichlids and comes in the second place of the world produced freshwater fishes (F.A.O., 2012, 2014). However, the input of aquaculture and tilapia to the total African production of animal protein, it is still inadequate (F.A.O., 2018, 2020). Boosting tilapia production may offer an improving strategy for adopting aquaculture to increase animal protein and raise Africa's currently very low protein intake. Tilapia is among the most extensively cultivated fish worldwide. Farmed tilapia accounts for more than 77% of global tilapia output (F.A.O., 2009).

Murphy et al. (2020) has reported an increase in the national aquaculture production, that review was prepared in the period from 2000 to 2016 and detected that increase starting 3.5×10^5 Mt toward 13.7×10^5 Mt. Tilapia particularly improved from almost 1.6×10^5 Mt in 2000 to 10×10^5 Mt in 2016. The Egyptian tilapia industry provides 47.9% of the total fish supply, which weighs around 941.308 Mt tilapia productions. Recently, the genetically improved *O. niloticus* was presented in Egypt, resulting in higher performance; with better-quality strains recording a 27.9 percent greater

harvest level (Ibrahim et al., 2013; Marjanovic et al., 2016 and Dickson et al., 2016).

According to estimates, Egypt ranks first and places in superior levels in aquaculture tilapia production compared to other African nations (Ali et al., 2020). Remarkably, the aquaculture industry secures around 585,000 jobs for workers in this area (El-Sayed et al., 2021). As a result, unanticipated deaths are thought to generate significant financial losses in aquaculture processes. It should be emphasized that the Egyptian Tilapia farms were mostly presented as small-scale farms of 10 feddan or less, accounting for approximately half of all *O. niloticus* farms (Eltholth et al., 2015).

2. Nile tilapia feeding behavior

Nile tilapia's dietary habits are herbivorous, even though it is an omnivore species. The farming of Tilapia is inexpensive because it nurses on a short chain of the feed in a fish farm, reducing the pressure on the species for preying. Furthermore, tilapia feeding patterns avoid the accumulation of contaminants at higher food chain levels (Barlow, 2000). Tilapia feed mostly on decomposing organic debris, and they can capture plankton in their gill mucus (Fryer and Iles, 1972). Tilapia has a lengthy gut that can digest planktons, and its feeding habits are classed as unceasing feeding

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action, which means they should have their daily meal on almost from 3 to 4 times. The important characteristic of *Oreochromis niloticus* is its sensitivity to cold weather, with an optimal level for development at 28 to 32 degrees Celsius. In the interim, a decrease of 31% in growth performance was reported with low temperature of 20 to 22°C (Lim, 1989).

3. Prebiotics

Prebiotics could be defined as any material, nutrient, substrate or fiber that assists as nutriment to the useful microbes in a host digestive tract (Mountzouris, 2022). Prebiotics should be able to withstand stomach acidity. Thus, a prebiotic could also be described as a material that resist acidic gastric conditions, might be fermented by gut microorganisms and effectively stimulate the gut microbiota growth to advance the gut well-being and host status (Davani-Davari *et al.*, 2019). They should be fermentable by the helpful gut microorganisms. Furthermore, prebiotics should encourage the beneficial microorganism's growth that benefits the host's health (Guerriero *et al.*, 2017). Prebiotics are famous for their favorable properties on gut micro-flora health and were reported to aid in the healing route of gut microorganisms following gut symbiosis in illness circumstances (Akram *et al.*, 2019). Prebiotics are hypothesized to provide health advantages to the gut by boosting the amount of gut micro-flora at the expense of dangerous microorganisms (Khangwal and Shukla, 2019). One method to overcome antibiotic overuse in aquatic health management is to employ prebiotics as feed additives (Song *et al.*, 2014; Zulkhisyam *et al.*, 2020; Kari *et al.*, 2021, 2022). These incorporations are being widely described to produce encouraging results in terms of growth enhancement (Ramos *et al.*, 2017), improving immunological reactions (Selim and Reda, 2015), resistance to various diseases (Li *et al.*, 2018), and relieving abiotic stressors in aquatic animals (Hoseinifar *et al.*, 2014).

3.1. Prebiotics in aquaculture

Prebiotics, often known as helpful compounds, offer food or energy to beneficial microbes. Prebiotics were used to improve fish health by favoring gut microbiota growth (Rohani *et al.*, 2021). Arabinoxylan oligosaccharide (AXOS), β -glucan, inulin, galacto-oligosaccharide (GOS), oligosaccharides, fructo-oligo saccharides (FOS), and mannan-oligosaccharide (MOS) were shown to improve aquaculture species' growth indicators (Li *et al.*, 2018), feed consumption efficacy (Shoaei *et al.*, 2015), immune response (Li *et al.*, 2005, Li *et al.*, 2021), and disease resistance (Figure, 1) (Yilmaz *et al.*, 2022). As a result, no doubt using prebiotics can aid to boost aquatic animals' output. Nevertheless, the fish gut biota is prebiotic-selective. Not all prebiotics are universal in their ability to encourage the expansion of the gut flora. Prebiotics like FOS (fructo-oligosaccharides) and GOS (galacto-oligosaccharides), for example, have been shown to enhance gut bacteria like *Bifido bacterium* and *Lactobacillus* sp. (Gibson *et al.*, 2017). This is due to the enzymes that *Bifido bacterium* and *Lactobacillus* sp. both have that ferment oligosaccharides. Regardless of the facts, more prebiotics added to fish feed can encourage more gut flora species of aquatic animals, improving their health (Teitelbaum and Walker, 2002). A Prebiotic can promote host gut flora growth (Dawood and Koshio, 2016). This might help in the development of aquatic creatures. Prebiotics can also improve feed efficacy (Figure 2). As a result, it can reduce feed conversion ratio (FCR) and boost aquatic animal growth performance. According to a literature review, numerous different forms

of prebiotic mixes efficiently support the growth of several aquatic species (Ganguly *et al.*, 2013). According to the literature review, several prebiotic mixtures successfully stimulate the growth of many fish and aquaculture species.

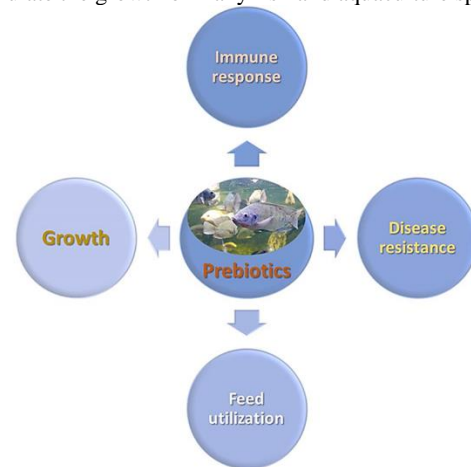


Figure 1 Prebiotics usage in aquaculture

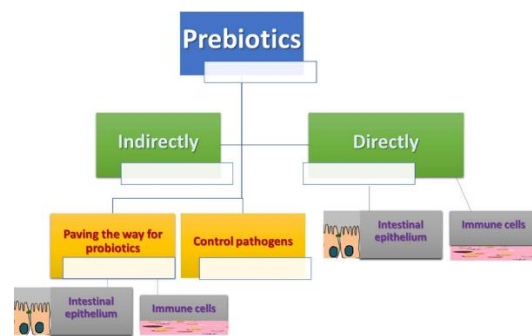


Figure 2 Prebiotics mode of action in aquaculture

3.1.1. Prebiotic supplementations effects on aquatic animals' growth performance

Gut microbiota could efficiently promote the growth of fish (Dawood and Koshio, 2016). This will assist in boosting the growth of fish and other aquatic animals. In addition, Prebiotics preparations can likewise upsurge feed usage and efficiency. Thus, it can raise the growth rate and minor the feed conversion rate (FCR) of aquatic animal (Ganguly *et al.*, 2013). It was reported that various mixtures of prebiotic and supplementations successfully promote the growth and weight gain of several aquatic species (Lu *et al.*, 2019). For instance, red drum (*Sciaenops ocellatus*) supplemented with GOS + FOS + Galacto-gluco-mannans (GGM) + MOS mixtures (Zhou *et al.*, 2010), post-larvae of *Litopenaeus vannamei* supplemented with GOS + inulin + FOS (Oktaviana and Yuhana, 2014), Chinese mitten crab (*Eriocheir sinensis*) incorporated with MOS + inulin + β -glucan (Lu *et al.*, 2019) and much more for example presented in Table 1. β -glucan prebiotic, was reported to improve the growth of many aquatic animals when used separately or in combination with several additional prebiotics. β -glucan, when combined with MOS will represent a perfect mixture that successfully can boost the Nile tilapia (*O. niloticus*) growth performance (Selim and Reda, 2015), and was reported to exert the same actions in other species for example, shabout (*Tor grypus*) (Mohammadian *et al.*, 2021), Chinese mitten crab (*Eriocheir sinensis*) (Lu *et al.*, 2019), snake head (*Channa striata*) (Munir *et al.*, 2016), and Caspian trout (*Salmo trutta caspius*) (Jami *et al.*, 2019).

Table 1 Effect of prebiotic supplementations on aquatic animals' growth performance.

Supplement	Fish spp.	Concentration g/kg diet	Reference
Glucans		0.1 for two weeks	(Lu <i>et al.</i> , 2019)
Mannan oligosaccharide and β -glucan	Nile tilapia	0.4-2	(Abdel gayed <i>et al.</i> , 2021)
Mannan oligosaccharide, β -glucan, and dextrose	<i>O. niloticus</i>	0.5–1.6	(Ismail <i>et al.</i> , 2019)
lecithin Lysomax [®]		0.45	(El-Sayed <i>et al.</i> , 2021)
β -glucan, MOS	Caspian trout	4	(Jami <i>et al.</i> , 2019)

Calcium format prebiotics when used separately or as a mixture with others at the rate of 0.5 and 1.5 percent can enhance the growth and feed conversion of fishes. Moreover, ACIDLAC[®] organic acids mixture when used as a replacement will have growth enhancement and other advantageous effects (Benedetto, 2003). Mairoka *et al.* (2004) similarly conveyed that organic acids compounds might successfully substitute antibiotic growth promoters (AGPs) and other chemicals for improved overall performance.

Almost synchronization occurs between probiotics and prebiotics, where probiotics augments the theory of prebiotics, through selectively stimulating the growth of gut beneficiary bacteria plus activating the metabolism of those bacteria that will improve the intestinal stability. However, the data about the prebiotics applications in aquaculture is still inadequate. It is reported that prebiotic Fermacto[®] addition (1, 2 and 3 g/kg) in aqua feeds displayed considerably higher body weight and better food conversion ratio in common carp fry. Rearing conditions of carp fry were improved after feeding Fermacto prebiotic at an optimal level of 3 g/kg (Mazurkiewicz *et al.*, 2008).

3.1.2. Effect of dietary phospholipid (PL) supplementation in aquafeeds

In larval and juvenile stages of several species of fish and crustaceans, dietary phospholipid (PL) supplementation in purified diets has been shown to positively affect survival, growth, resistance to stress testing, and abnormalities. Due to the difficulties of bio-encapsulating PL in live prey, it is difficult to determine the precise PL needs in larvae. It is also challenging to compare needs estimated with artificial diets due to the wide variation in purity and content of PL sources and experimental settings (such as diet formulation and degree of co/pre-feeding with real food). Larval stages need more nutritional PL than juveniles because they are so vulnerable to a dietary PL shortage. Red sea bream larvae and goldfish larvae both had substantial reductions in survival. As evidenced by the survival of young fish, such as rock bream *Oplegnathus fasciatus* and sea bass, juvenile fish appear to be less impacted by the dietary PL supply than larvae (Kanazawa, 1993).

There is substantial debate about whether high dietary phospholipid levels are harmful to fish and crustaceans. Increasing dietary PL levels over the necessary level did not affect survival or growth in several trials (Kanazawa, 1993). However, when more than 3% soybean lecithin was included in the diet, larval *P. japonicus* survival was reduced (Coutteau *et al.*, 1996).

It has been proposed that lecithin is needed as a surfactant for effective lipid emulsification and digestion in early crustacean and fish larval stages (Koven *et al.*, 1998). Dietary PL is more important in larval fish and crustaceans than in juveniles. Because of their capacity to transport lipids and retain fatty acids, PL fractions such as Phosphatidyl choline (PC) and PI are more effective in larvae. Furthermore, PLs increase food features regarding water

stability via their gum characteristics and chemo-attractive capabilities. Because of its increased digestibility, it outperforms neutral lipids in terms of essential fatty acid and energy content. The favorable effects of PL lead to increased development, survival, and quality in fish and crustacean larvae. Sea bass larvae can use PL more effectively than neutral lipids, resulting in increased growth, survival, and skeletal development Cahu *et al.*, 2003; Olsen *et al.*, 2014). The key bottleneck is the reduced survival of freshwater catfish (*Clarius batrachus* and *Heteropneustus fossilis*) larvae, which might be addressed by PL supplementation in the diet (Olsen *et al.*, 2014). As a result, further studies on these species are required.

3.1.3. Effects of prebiotic mixtures containing soybean lecithin on the growth performance of aquatic animals

The fish performance and innate immune response can be significantly enhanced by using the lecithin-containing bio-emulsifier Lysomax[®] for farmed Nile tilapia to perform at their optimum and maintain good health, around 0.45 g/kg of feed would be sufficient (El-Sayed *et al.*, 2021).

The aquafeed industry has been obliged to lower the quantities of dietary fish oil in feed formulations due to the constant rise in fish oil costs and their unsustainable supply. An extensive study has been done on partially substituting plant oil with fish oil (Santigosa *et al.*, 2011).

Freshwater fish respond differently to additional PLs depending on the species, stage of growth, and environmental factors. In larval common carp (*Cyprinus carpio*), for instance, dietary PL was crucial for growth, survival, feed utilization, and optimal health, most likely because immature fish cannot synthesis PL (Fontagne *et al.*, 2000). However, because they can produce PL from nutritional precursors, bigger fish may not benefit from more PL. Supplemental soy lecithin, on the other hand, had no impact on fingerling and juvenile channel catfish survival, growth rates, or feed efficiency (Sink and Lochmann, 2014).

3.1.4. Prebiotic supplementations effect on aquatic animals' immune response and disease resistance

Prebiotic could be an intricate carbohydrate that might offer a source for energy for gut macrobiotic abundance, therefore it could improve the immune response of aquatic animals and fishes (Mohammadi *et al.*, 2022). Prebiotics can motivate immune reactions in fishes. The immune saccharides might be an additional selection to be administered in aquaculture as an alternative to the usage of antibiotics for enhancing the health management of aquatic animals (Hoseinifar *et al.*, 2020). There are two expending methods for the prebiotics to improve the immune reactions in aquatic animals, whichever through demotion of gut flora growth or innate immune system activation (Song *et al.*, 2014). Nevertheless, the microbiota of animal gut is selective to an exact prebiotic. Thus, the prebiotic mixtures application will provide superior effects on the immune response of aquatic animals (Elumalai *et al.*, 2022) as revealed in Table 2.

Table 2 Effect of probiotic supplementation on the aquatic animals' immune response.

Supplement	Species	Dose g/kg diet	Reference
Chito-oligosaccharides and Shrimp shell chitin (hydrolyzed)	Tilapia hybrid (<i>O.aureus</i> × <i>O.niloticus</i>)	1	(Qin <i>et al.</i> , 2014)
GOS + FOS + GGM+ MOS	Red drum	10	(Zhou <i>et al.</i> , 2010)
MOS and β-glucan	Caspian trout	4	(Jami <i>et al.</i> , 2019)
	Gilthead sea bream	1	(Guzmán-Villanueva <i>et al.</i> , 2014)
	Nile tilapia	0.5-2	(Abdel gayed <i>et al.</i> , 2021)

There are very few studies on immune modulation by dietary PL. Phosphatidyl serine has the immune modulatory role in white shrimp (*Penaeus vannamei*) when supplemented in diet that can enhance total hemocyte count and phenoloxidase values of hemocyte and plasma. Besides these, it can also modulate antibacterial and bacteriolytic activities in blood plasma, and PL can modulate anti-stress capacity in shellfish. Inclusion of PL in diet improves the gill health of grass carp, indicating enhanced immunity and antioxidant status and it is also reported that supplementation of PL in diet can improve gut health and immunity through modulation of lysozyme, up-regulating expression level of interleukin 10, down-regulated tumor necrosis factor α (TNF-α), superoxide dismutase, glutathione peroxidase and glutathione reductase (Cui *et al.*, 2018).

Similar to this, immune responses (phagocytic activity, respiratory burst activity lysozyme activity, and phenoloxidase values) in *O. niloticus* fed lecithin-containing diets, particularly at 0.4 g/kg, were greatly improved compared to the control diet-fed fishes. These findings unequivocally show that Lysomax enhances innate immunity in this species. This may be explained by the fact that dietary PLs and their unsaturated fatty acids can make cell membranes more permeable and fluid, which strengthens fish immune (Balfry and Higgs, 2001).

The increased phospholipid composition and fatty acid outline of Lysomax may also be responsible for improving fish immunity as a result of supplementation. Phospholipids are the precursor of the substrate fatty acids needed for producing eicosanoids (EPA, 20:5n-3) (Balfry and Higgs, 2001). Most of the research highlighted in this review focused on the favorable benefits of probiotic combinations as feed additives, which can boost aquaculture output by encouraging growth, strengthening the immune system, raising disease resistance, and lowering stress in aquatic animals. Research has been undertaken to assess the potential of probiotics or probiotic combinations to lower stress in aquatic animals. However, when introducing probiotics to aquaculture species, correct dose and probiotic type considerations must be made. A high dose of probiotics can be harmful to aquatic animals, whilst other types of probiotics can only boost growth but not the immune response or resistance to diseases of aquatic animals. There is still a gap between practical applications and scientific findings of probiotics in aquatic animals.

REFERENCES

- Abdel Gayed, M., Abbass, A., Shehabeldin, M., Elabd, H., 2021, 'Fermos® probiotic dietary supplementation enhances immune, antioxidative responses and growth performance of Nile tilapia *Oreochromis niloticus*', Benha Veterinary Medical Journal, 40(1), 135-140.
- Akram, W., Garud, N., Joshi R., 2019, 'Role of inulin as probiotics on inflammatory bowel disease', Drug Discoveries & Therapeutics, 2019, 13(1), 1-8, Released on J-STAGE Online ISSN 1881-784X, Print ISSN 1881-7831
- Ali SE, Jansen MD, Mohan CV, Delamare-Deboutteville, J., and Charo-Karisa, H., 2020, 'Key risk factors, farming practices and economic losses associated with tilapia mortality in Egypt', Aquaculture 527, 735438. <https://dx.doi.org/10.1016/j.aquaculture.2020.735438>.
- Balfry, S. K., and Higgs, D. A., 2001, 'Influence of dietary lipid composition on the immune system and disease resistance of finfish', Nutrition and Fish Health, 12, 213–234.
- Barlow, GW., 2000, 'The Cichlid fish. Nature's grand experiment in evolution Perceous books group Cambridge', Massachusetts ISBN0-7382-0376-9.
- Benedetto, M., 2003, 'The effect of organic acids (Acid Lac® Reg. Micro PELLETS) on breeder and on gut health maintenance', Zoo tecnica Int 6, 40–45
- Cahu, C. L., Zambonini, I. J., and Barbosa, V., 2003, 'Effect of dietary phospholipid and phospholipid: neutral lipid value on the development of sea bass (*Decentrarchus labrax*) larvae fed a compound diet', Br. J. Nutr., 90, 21–28.
- Coutteau, P., Camara, M. R., and Sorgeloos, P., 1996, 'The effect of different levels and sources of dietary phosphatidyl choline on the growth, survival, stress resistance, and fatty acid composition of post larval *Penaeus vannamei*', Aquaculture, 147, 261-273.
- Cui, X., Zhang, Qi., Zhang, Qu., Zhang, Yo., Chen, H., Liu, G., and Zhu, L., 2022, 'Research Progress of the Gut Microbiome in Hybrid Fish', Microorganisms, 10, (5), 891. doi.org/10.3390/microorganisms10050891
- Davani-Davari, Dorna, Manica Negahdaripour, Iman Karimzadeh, Mostafa Seifan, Milad Mohkam, Seyed Jalil Masoumi, Aydin Berenjian, and Younes Ghasemi, 2019, 'Prebiotics: Definition, Types, Sources, Mechanisms, and Clinical Applications', Foods, 8(3), 92.
- Dawood, M.A.O., and Koshio, S., 2016, 'Recent advances in the role of probiotics and prebiotics in carp aquaculture', A review. Aquaculture journal, 454, 243-251.
- Dickson, M., Nasr-Allah, A., Kenawy, D., and Kruijssen, F., 2016, 'Increasing fish farm profitability through aquaculture best management practice training in Egypt', Aquaculture, 465, 172-178.
- El-Sayed, A. F.M., Tammam, M.S. and Makled, S.O., 2021, 'Lecithin-containing bioemulsifier boosts growth performance, feed digestion and absorption and immune response of adult Nile tilapia (*Oreochromis niloticus*)', Aquacult, Nutr, 27, (3), 757-770.
- Elsaied, H.E., Soliman, T., Abu-Taleb, H.T., Goto, H., and Jenke-Kodam, H., 2019, 'Phylogenetic characterization of eukaryotic and prokaryotic gut flora of Nile tilapia, *Oreochromis niloticus*, along niches of lake Nasser, Egypt, based on rRNA gene high-throughput sequences', Ecol Genet Genom, 11, 100037. <https://doi.org/10.1016/j.egg.2019.100037>.
- Eltholth, M., Forance, K., Grace, D., Rushton, J., and Hasler, B., 2015, 'Characterization of production, marketing and consumption patterns of farmed tilapia in the Nile delta of Egypt', Food Policy, 51, 131-143.
- Elumalai, P., Kurian, A., Lakshmi, S., Mustafa, M. S., Ring, E., and Faggio, C., 2022, 'Effect of *Leucospora* against *Aeromonas hydrophila* in Nile Tilapia (*Oreochromis niloticus*), Immunity and gene expression evaluation' Turk, J, Fish & Aquat, Sci, 22, (2), 19802.
- F.A.O., 2009, 'Information and knowledge sharing Food and Agriculture Organization of the United Nations Technical Guidelines for Responsible Fisheries', (12), Rome, 97p.
- F.A.O., 2012, 'Food and Agriculture Organization of the United Nations', Accessed July 20, 2012 .
- F.A.O., 2014, 'Management of inland waters for fish: a cross-sectoral and multi-disciplinary approach. In The state of world

- fisheries and aquaculture 2014', Food and Agriculture Organization of the United Nations, Rome, 116–120.
20. F.A.O., 2018, 'Licence C, (Ed.), state of fisheries and aquaculture in the world 2018-meeting in the sustainable development goals', Food and agriculture organization of United Nations, Rome, 227 p.
 21. F.A.O., 2020, 'The State of World Fisheries and Aquaculture 2020', Sustainability in action, Rome.
 22. Fontagne, S., Burtaire, L., Corraze, G., and Bergot, P., 2000, 'Effects of dietary medium-chain triacylglycerols (tricaprylin and tricaproin) and phospholipid supply on survival, growth and lipid metabolism in common carp (*Cyprinus carpio* L.) larvae', *Aquaculture*, 190, 289–303.
 23. Fryer, G., and Iles, T.D., 1972 'The Cichlid fish of the great lakes of Africa: Their biology and evolution', TFH Publications, Hong Kong, 641.
 24. Ganguly, S., Dora, K. C., Sarkar, S., and Chowdhury, S., 2013, 'Supplementation of prebiotics in fish feed: A review', *Reviews in Fish Biology and Fisheries*, 23, (2), 195–199.
 25. Gibson, G. R., Hutkins, R., Sanders, M. E., Prescott, S. L., Reimer, R. A., Salminen, S. J., Scott, K., Stanton, C., Swanson, K. S., and Cani, P. D., 2017, 'Expert consensus document: The International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics', *Nature Reviews Gastroenterology and Hepatology*, 14, (8), 491–502.
 26. Guerriero, I., Oliva-Teles, A., and Enes, P., 2017, 'Prebiotics as functional ingredients: Focus on Mediterranean fish aquaculture', *Rev Aquaculture*, 0, 1-33.
 27. Guzmán Villanueva, L.T., D., Tova, Ramírez, E., Gisbert, H., Cordero, F.A., Guardiola, A., Cuesta, J., Meseguer, F., Ascencio-Valle, M.A., Esteban, 2014, 'Dietary administration of β -1,3/1,6-glucan and probiotic strain *Shewanella putrefaciens*, single or combined, on gilthead seabream growth, immune responses and gene expression', *Fish & Shellfish Immunology*, 39, (1), 34-41.
 28. Hoseinifar, S. H., Soleimani, N., and Ringø, E., 2014, 'Effects of dietary fructo-oligosaccharide supplementation on the growth performance, haemato-immunological parameters, gut microbiota and stress resistance of common carp (*Cyprinus carpio*) fry', *British Journal of Nutrition*, 112, (8), 1296–1302.
 29. Ibrahim, N.A., Zaid, M.Y.A., Khaw, H.L., El-Naggar, G.O., and Ponzoni, R.W., 2013, 'Relative performance of two Nile tilapia (*Oreochromis niloticus* Linnaeus) strains in Egypt: The Abbassa selection line and the Kafr Elsheikh commercial strain', *Aquac. Res.*, 44, 508-517.
 30. Ismail, M., A., Wahdan, M.S., Yusuf, E., Metwally, M., Mabrok, 2019, 'Effect of dietary supplementation with a synbiotic (Lacto Forte) on growth performance, hematological and histological profiles, the innate immune response and resistance to bacterial disease in *Oreochromis niloticus*', *Aquaculture Research*, 50, (9), 2545-2562.
 31. Jami M.J., A.A., Kenari, H., Paknejad, M., Mohseni, 2019, 'Effects of dietary β -glucan, mannan oligosaccharide, *Lactobacillus plantarum* and their combinations on growth performance, immunity and immune related gene expression of Caspian trout, *Salmo trutta caspius* (Kessler, 1877)', *Fish & Shellfish Immunology*, 91, 202-208.
 32. Kari, Z. A., Kabir, M. A., Dawood, M. A., Razab, M. K. A. A., Ariff, N. S. N. A., Sarkar, T., Pati, S., Edinur, H. A., Mat, K., Ismail, T. A., and Wei, L. S., 2022, 'Effect of fish meal substitution with fermented soy pulp on growth performance, digestive enzyme, amino acid profile and immune-related gene expression of African catfish (*Clarias gariepinus*)', *Aquaculture*, 546, 737418.
 33. Kari Abdul, Z., Kabir, M. A., Mat, K., Rusli, N. D., Razab, M. K. A. A., Ariff, N. S. N. A., Edinur, H. A., Rahim, M. Z. A., Pati, S., Dawood, M. A., and Wei, L. S., 2021, 'The possibility of replacing fish meal with fermented soy pulp on the growth performance, blood biochemistry, liver, and intestinal morphology of African catfish (*Clarias gariepinus*)', *Aquaculture Reports*, 21, 100815. <https://doi.org/10.1016/j.aqrep.2021.100815>.
 34. Kanazawa, A., 1993, 'Essential phospholipids of Fish and crustaceans', In *fish Nutrition in practice* (eds. Kaushik S J and Luquet P), edition INRA les colloques (61), 519-530, Biarrit, France.
 35. Khangwal, I., and Shukla, P., 2019, 'Potential prebiotics and their transmission mechanisms: recent approaches', *JFDA*, 27, 649-656.
 36. Koven, W. M., Kolkovski, S., and Tandler, A., 1998, 'The effect of dietary phosphatidylcholine and its constituent fatty acids on microdiet ingestion and fatty acid absorption rate in gilthead sea bream, *Sparus auratus*, larvae', *Aquaculture Nutrition*, 4, 39–45.
 37. Li, P., and Gatlin, D.M., I. I.I., 2005, 'Evaluation of the prebiotic GroBiotic-A and brewer's yeast as dietary supplements for sub-adult hybrid striped bass (*Morone chrysops* x *M. saxatilis*) challenged in situ with *Mycobacterium marinum*', *Aquaculture*, 248, 197-205.
 38. Li, Y., Liu, H., Dai, X., Li, J., and Ding, F., 2018, 'Effects of dietary inulin and mannan oligosaccharide on immune related genes expression and disease resistance of pacific white shrimp', *Fish and shellfish immunology*, 76, 78-92.
 39. Li, Y., Yuan, W., Zhang, Y. et al., 2021, 'Single or combined effects of dietary arabinoxylan-oligosaccharide and inulin on growth performance, gut microbiota, and immune response in Pacific white shrimp *Litopenaeus vannamei*', *J. Ocean, Limnol*, 39, 741–754.
 40. Lim, C., 1989, 'Practical Feeds-Tilapia, in: RT Lovell (ed.), *Nutrition and feeding of fish*', (Van Nostrand Reinhold, New York, New York, USA), 163-167.
 41. Lu, J., Bu, X., Xiao, S., Lin, Z., Wang, X., Jia, Y., Wang, X., Qin, J. G., and Chen, L., 2019, 'Effect of single and combined immunostimulants on growth, anti-oxidation activity, non-specific immunity and resistance to *Aeromonas hydrophila* in Chinese mitten crab (*Eriocheirsinensis*)', *Fish and Shellfish Immunology*, 93, 732–742.
 42. Mairoka, A., Santin, A. M. E., Borges, S.A., Opalinski, M., Silva, A.V.F., 2004, 'Evaluation of a mix of fumaric, lactic, citric and ascorbic acids on starter diets of broiler', *Arch Vet Sci*, 9, (1), 31–37.
 43. Marjanovic, J., Mulder, H. A., Khaw, H. L., and Bijma, P., 2016, 'Genetic parameters of uniformity of harvest weight and body size traits in the GIFT strain of Nile tilapia, *Genet. Sel. Envol*, 48, 41 <https://doi.org/10.1186/s12711-016-0218-9>.
 44. Mazurkiewicz, J., Przybył, A., Golski, J., 2008, 'Usability of FERMacto prebiotic in feeds for common carp (*Cyprinus carpio* L.) fry', *NaukaPrzyrTechnol*, 2, (3), 15.
 45. Mohammadi, G., Hafezieh, M., Karimi, A. A., Azra, M. N., Doan, H. V., Tapingkae, W., Abdelrahman, H. A., and Dawood, M. A. O., 2022, 'The synergistic effects of plant polysaccharide and *Pediococcus acidilactici* as a synbiotic additive on growth, antioxidant status, immune response, and resistance of Nile tilapia (*Oreochromis niloticus*) against *Aeromonas hydrophila*', *Fish and Shellfish Immunology*, 120, 304–313.
 46. Mohammadian, T., Ghanei Motlagh, Molayemraftar, M., Mesbah, M., Zarea, Mohtashampour, A.J., Nejad, 2021, 'Modulation of growth performance, gut microflora, non-specific immunity and gene expression of proinflammatory cytokines in shabout (*Tor grypus*) upon dietary prebiotic supplementation', *Fish & Shellfish Immunology*, 112, 38-45.
 47. Mountzouris, K. C., 2022, 'Prebiotics: Types', *Encyclopedia of dairy sciences*, 3rd ed., 352–358.
 48. Munir, M.B., R., Hashim, Y.H., Chai, T.L., Marsh, S.A.M., Nor 2016, 'Dietary prebiotics and probiotics influence growth performance, nutrient digestibility and the expression of immune regulatory genes in snakehead (*Channa striata*) fingerlings', *Aquaculture*, 460, 59-68.
 49. Murphy, S., Charo-Karisa, H., Rajaratnam, S., Cole, S. M., McDougal, C., Nasr-Allah, A. M., Kenawy, D., AbouZead, M. Y., van Brakel, M. L., Banks, L. K., and Ibrahim, N., 2020, 'Selective breeding trait preferences for farmed tilapia among low-income women and men consumers in Egypt: implications for poor-poor and gender-responsive fish breeding programs', *Aquaculture*, 525, 735042.
 50. Nawaz, A., Irshad, S., Hoseinifar, S. H., and Xiong, H., 2018, 'The functionality of prebiotics as immunostimulant: Evidence

- from trials on terrestrial and aquatic animals', *Fish and Shellfish Immunology*, 76, 272–278.
51. Oktaviana, A., and Yuhana, M., 2014. The use of synbiotics to prevent *imnv* and *Vibrio harveyi* co-infection in *Litopenaeus vannamei*', *HAYATI Journal of Biosciences*, 21, (3), 127–134.
 52. Olsen, Y., Evjemo, J.O., Kjørsvik, E., Larssen, H., Li, K., Overrein, I., Rainuzzo, J., 2014. DHA content in dietary phospholipids affects DHA content in phospholipids of cod larvae and larval performance. *Aquaculture* 428–429, 203–214.
 53. Qin, C., Y., Zhang, W., Liu, L., Xu, Y., Yang, Z., Zhou, 2014. Effects of chito-oligosaccharides supplementation on growth performance, intestinal cytokine expression, autochthonous gut bacteria and disease resistance in hybrid tilapia *Oreochromis niloticus*♀× *Oreochromis aureus*♂', *Fish & Shellfish Immunology*, 40, (1), 267-274.
 54. Ramos, M. A., Batista, S., Pires, M. A., Silva, A. P., Pereira, L. F., Saavedra, M. J., Ozório, R. O. A., and Rema, P., 2017. 'Dietary probiotic supplementation improves growth and the intestinal morphology of Nile tilapia', *Animal*, 11, 1259–1269.
 55. Rohani, M. F., Islam, S. M., Hossain, M. K., Ferdous, Z., Siddik, M. A., Nuruzzaman, M., Padeniya, U., Brown, C., and Shahjahan, M., 2021. 'Probiotics, prebiotics and synbiotics improved the functionality of aquafeed: Upgrading growth, reproduction, immunity and disease resistance in fish', *Fish and Shellfish Immunology*, 120, 569–589.
 56. Santigosa, E., García-Meilán, I., Valentín, J. M., Navarro, I., Pérez- Sánchez, J., and Gallardo, M. Á., 2011. 'Plant oils' inclusion in high fish meal-substituted diets: effect on digestion and nutrient absorption in gilthead sea bream (*Sparus aurata* L.)', *Aquaculture Research*, 42, 962–974.
 57. Selim, K. M., and Reda, R. M., 2015. 'Beta-glucans and mannan oligosaccharides enhance growth and immunity in Nile tilapia', *North American Journal of Aquaculture*, 77, (1), 22–30.
 58. Shoaee, R., Akrami, R., Ghobadi, S., and Razeqhi Mansour, M., 2015. 'Effect of dietary of prebiotic mannan oligosaccharide and β -1, 3 glucan on growth performance, survival, body composition and serum lysozyme activity in Rainbow trout (*Oncorhynchus mykiss*) fingerling', *Journal of Marine Biology*, 7, (2), 45–56.
 59. Sink, T. D., and Lochmann, R. T., 2014. 'The effects of soybean lecithin supplementation to a practical diet formulation on juvenile channel catfish, *Ictalurus punctatus*: growth, survival, hematology, innate immune activity, and lipid biochemistry', *Journal of the World Aquaculture Society*, 45, 163– 172 .
 60. Song, S. K., Beck, B. R., Kim, D., Park, J., Kim, J., Kim, H. D., & Ringø, E., 2014. 'Prebiotics as immunostimulants in aquaculture: A review', *Fish & Shellfish Immunology*, 40, (1), 40–48. <https://doi.org/10.1016/j.fsi.2014.06.016>.
 61. Teitelbaum, J. E., and Walker, W. A., 2002. 'Nutritional impact of pre-and probiotics as protective gastrointestinal organisms', *Annual Review of Nutrition*, 22, (1), 107–138.
 62. Yılmaz, S., Yılmaz, E., Dawood, M. A., Ringø, E., Ahmadifar, E., and Abdel-Latif, H. M. R., 2022. 'Probiotics, prebiotics, and synbiotics used to control vibriosis in fish: A review', *Aquaculture*, 547, Article 737514 .
 63. Zhou, Q.C., Buentello, J. A., and Gatlin, D. M., I.I.I., 2010. 'Effects of dietary prebiotics on growth performance, immune response and intestinal morphology of red drum (*Sciaenops cellatus*)', *Aquaculture*, 309, (1–4), 253–257.
 64. Zulhisyam, A. K., Kabir, M. A., Munir, M. B., and Wei, L. S., 2020. 'Using of fermented soy pulp as an edible coating material on fish feed pellet in African catfish (*Clarias gariepinus*) production', *Aquaculture, Aquarium, Conservation and Legislation*, 13, (1), 296–308.