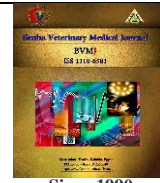




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### Original Paper

## Occurrence of anaerobic spore formers including *Clostridium perfringens* and its virulence genes in some vacuum-packed chilled meat products

Alaa Abd El-Satar<sup>1,2</sup>, Saad, M. Saad<sup>1</sup>, Nahla A. Abo EL-Roos<sup>2</sup>, Mohamed A. Hassan<sup>1</sup>

<sup>1</sup>Department of Food control, Faculty of Veterinary Medicine, Benha University, Egypt

<sup>2</sup>Department of Food Hygiene, Animal Health Research Institute, Shebin El-Kom branch, Agriculture Research Center, Egypt

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### ABSTRACT

Ninety samples of various vacuum-packed meat products, including sausage, frankfurter and salami, were selected randomly from several Menofya governorate supermarkets in order to be examined for the presence of *Clostridium perfringens* and its virulence genes, as well as anaerobic spore formers. The results showed that the prevalence of anaerobic spore forming bacteria was 18 (60%), 25(83.33%) and 26 (86.67%) in salami, frankfurter and sausage, respectively. The mean values of anaerobic spore forming bacteria count in vacuum packed salami, frankfurter and sausage were  $1.09 \times 10^3 \pm 0.17 \times 10^3$ ,  $3.22 \times 10^3 \pm 0.58 \times 10^3$  and  $5.61 \times 10^3 \pm 1.20 \times 10^3$ , respectively. Also, the prevalence of *C. perfringens* in same products was 46.67%, 63.33% and 70%, respectively. Furthermore, the mean values of *C. perfringens* counts for salami, frankfurter and sausage were  $9.95 \times 10^2 \pm 2.14 \times 10^2$ ,  $2.18 \times 10^3 \pm 0.37 \times 10^3$  and  $3.87 \times 10^3 \pm 0.65 \times 10^3$  CFU/g, respectively. The acceptability of same products based on their contamination with *C. perfringens*, was 53.33%, 36.67% and 30 % was accepted, while 46.67%, 63.33% and 70% were unaccepted, respectively. Moreover, *cpa*(402 bp), *cpb* (236 bp), *cpx* (541bp) and *iap* (317 bp) *C. perfringens* virulence enterotoxin genes was detected from meat products by using multiplex PCR and their incidence were 50%, 50% and 25% for *cpa* gene while it was 25%, 25% and 25 % for *cpa* and *cpb* genes and 25%, 25% and 25% for *cpa* and *cpx* genes and 0%, 0% and 25% for *cpa*, *cpb* and *iap* genes isolated from the salami, frankfurter and sausage meat products respectively. Sausage samples were the most contaminated products. So, periodical examination and HACCP system application considered very important in meat products plants.

## 1. INTRODUCTION

Meat processing sector frequently encounters numerous significant issues pertaining to the safety and hygiene of diverse products (Ruiz-Capillas and Herrero, 2019). Pathogen transmission into the product may occur from the carcass surface or during handling procedures (Palmer et al., 2007). Anaerobic spore-forming foodborne pathogen as *Clostridium perfringens* is found in a vast range of foods, including meat and fish products. Because *C. perfringens* spores are heat resistant, they may withstand the heating process, germinate, and spread throughout food (Chang et al., 2020). This pathogen produces diarrheal illness (Limbo et al., 2010) or even mortality (Grass, et al. 2013) by affecting the permeability of the cell membrane in the intestine when the contaminated food is consumed. Foodborne infections have been linked to multiple outbreaks and have been linked to a number of serious illnesses and public health issues (Okeke et al., 2005). Foodborne epidemics caused by *C. perfringens* were recently discovered during the Panhellenic Handball Championship for kids (Mellou et al., 2019) consequently, techniques for managing *C. perfringens*, particularly its spores in diverse meat products, are required (Juneja et al., 2013). When *C. perfringens* spores germinate in food and survive cooking, they can cause an outbreak of food poisoning. These germinated spores eventually cause enterotoxins to be

produced in the digestive tract following ingestion. Thus, the current study sheds insight on the number of anaerobic spore formers, particularly *C. perfringens* bacteria; found in sausage, frankfurters and salami as well as the multiplex PCR method used to identify their virulence genes.

## 2. MATERIAL AND METHODS

### 2.1. Collection of samples

Randomly selected ninety samples of vacuum-packed meat products, sausage, frankfurter, and salami 30 of each were gathered from various supermarkets in the Menofya governorate, Egypt, in order to look for anaerobic spore formers, specifically *C. perfringens* and its genes virulence

### 2.2. Preparation of samples

The collected samples were prepared according to APHA (2001).

### 2.3. Enumeration of anaerobic spore formers (ISO 15213:2003)

### 2.4. Enumeration and isolation of *C. perfringens*

The volume of 0.1 ml was pipetted into the center of tryptose sulphate cycloserine agar plates (TSC). The plates were incubated upright and anaerobically at 37°C for 48 hrs. The plates containing characteristic black colonies (1-2 mm in

diameter surrounded by district wide zone of opaque precipitate, 3-4 mm in diameter) were selected; the black colonies were counted (Collee et al., 1996).

2.5. Identification of *C. perfringens* (Koneman et al., 1992)

2.6. Detection of Virulence genes of *Clostridium perfringens* by PCR (Meer and Songer, 1997)

2.7. Statistical Analysis

ANOVA test was used to statistically examine the results (Feldman et al., 2003)

3. RESULTS

The results showed that the prevalence of anaerobic spore forming bacteria was 18(60%), 25(83.33%) and 26 (86.67%) in salami, frankfurter and sausage, respectively (table 1). Also, the mean counts of anaerobic spore forming bacteria of vacuum packed salami, Frankfurter and Sausage were  $1.09 \times 10^3 \pm 0.17 \times 10^3$ ,  $3.22 \times 10^3 \pm 0.58 \times 10^3$  and  $5.61 \times 10^3 \pm 1.20 \times 10^3$ , respectively (table 2).

Table (3) reported that the prevalence of *C. perfringens* in the vacuum packed Salami, Frankfurter and Sausage were 46.67%, 63.33% and 70%, respectively. Further, the mean values of *C. perfringens* count for Salami, frankfurter and sausage were  $9.95 \times 10^2 \pm 2.14 \times 10^2$ ,  $2.18 \times 10^3 \pm 0.37 \times 10^3$  and  $3.87 \times 10^3 \pm 0.65 \times 10^3$  CFU/g, respectively (table 4).

In table (5) the acceptability of vacuum packed Salami, Frankfurter and Sausage meat products based on their contamination with *C. perfringens* was 53.33%, 36.67% and 30% were accepted, while 46.67%, 63.33% and 70% were unaccepted, respectively. Moreover, in table (6) the incidence of virulence genes of *C. perfringens* were 50%, 50% and 25% for *cpa*, 25%, 25% and 25% for *cpa* and *cpb* and 25%, 25% and 25% for *cpa* and *cpx* and 0%, 0% and 25% *cpa*, *cpb* and *iap* isolated for Salami, Frankfurter and Sausage meat products respectively.

Figure (1) proved the presence of *cpa*(402 bp), *cpb* (236 bp), *cpx* (541bp) and *iap* (317 bp) virulence enterotoxin genes of *C. perfringens* that isolated from meat products by using Agarose gel electrophoresis of multiplex PCR

Table (1): Prevalence of anaerobic spore formers in the examined samples of vacuum packed meat products (n=30)

Meat products	No of positive samples	%
Salami	18	60
Frankfurter	25	83.33
Sausage	26	86.67
Total	69	76.67

Table (2): Statistics of anaerobic spore formers counts in the examined samples of vacuum packed meat products (n=30)

Meat products	Min	Min	Mean ± S.E <sup>†</sup>
Salami	$1.0 \times 10^2$	$4.7 \times 10^3$	$1.09 \times 10^3 \pm 0.17 \times 10^3$ <sup>a</sup>
Frankfurter	$1.0 \times 10^2$	$9.2 \times 10^3$	$3.22 \times 10^3 \pm 0.58 \times 10^3$ <sup>b</sup>
Sausage	$1.0 \times 10^2$	$1.5 \times 10^4$	$5.61 \times 10^3 \pm 1.20 \times 10^3$ <sup>c</sup>

S.E<sup>†</sup> = standard error of mean, the different superscripted small letters represent a the significance difference at level p ≤ 0.05

Table (3): Acceptability of vacuum packed meat products based on their contamination with anaerobic spore formers (n=30)

Meat products	Anaerobic count /g*	Accepted		Unaccepted	
		No	%	No	%
Salami	≤ 100	21	70	9	30
Frankfurter	≤ 100	13	43.3	17	56.7
Sausage	≤ 100	12	40	18	60
Total (90)		46	51.1	44	48.9

\*Egyptian Organization for Standardization "EOS" (2005)

Table (4): Prevalence of *Clostridium perfringens* in the examined samples of vacuum packed meat products (n=30)

Meat products	No of positive samples	%
Salami	14	46.7
Frankfurter	19	63.3
Sausage	21	70
Total	54	60

Table (5): Statistics of *Clostridium perfringens* counts in the examined samples of vacuum packed meat products (n=30)

Meat products	Min	Min	Mean ± S.E <sup>†</sup>
Salami	$1.0 \times 10^2$	$3.1 \times 10^3$	$9.95 \times 10^2 \pm 2.14 \times 10^2$ <sup>a</sup>
Frankfurter	$1.0 \times 10^2$	$6.0 \times 10^3$	$2.18 \times 10^3 \pm 0.37 \times 10^3$ <sup>b</sup>
Sausage	$1.0 \times 10^2$	$8.9 \times 10^4$	$3.87 \times 10^3 \pm 0.65 \times 10^3$ <sup>c</sup>

S.E<sup>†</sup> = standard error of mean, the different superscripted small letters represent a the significance difference at level p ≤ 0.05

Table (6): Acceptability of vacuum packed meat products based on their contamination with *Clostridium perfringens* (n=30)

Meat products	<i>Clostridium perfringens</i> count /g*	Accepted		Unaccepted	
		No	%	No	%
Salami	Free	16	53.3	14	46.7
Frankfurter	Free	11	36.7	19	63.3
Sausage	Free	9	30	21	70
Total (90)		36	40	54	60

\*Egyptian Organization for Standardization "EOS" (2005)

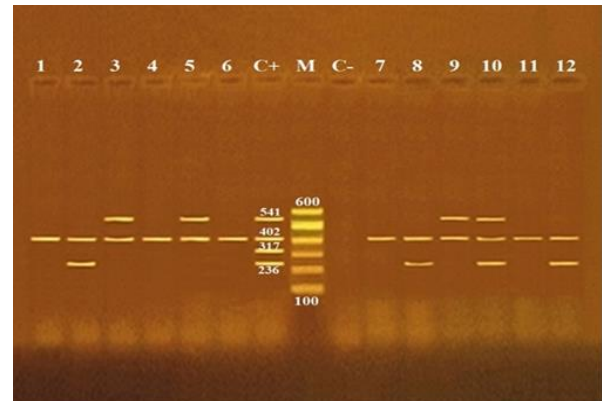


Figure (1): Agarose gel electrophoresis of multiplex PCR of *cpa* (402 bp), *cpb* (236 bp), *cpx* (541bp) and *iap* (317 bp) virulence enterotoxin genes of *C. perfringens*. Lane M: 100 bp ladder as molecular size DNA marker. Lane C+: Control positive for *cpa*, *cpb*, *cpx* and *iap* genes. Lane C-: Control negative. Lanes 1, 4, 6, 7 and 11: Positive *C. perfringens* strains for *cpa* gene. Lanes 2, 8 and 12: Positive *C. perfringens* strains for *cpa* and *cpb* genes. Lanes 3, 5 and 9: Positive *C. perfringens* strains for *cpa* and *cpx* genes. Lane 10: Positive *C. perfringens* strain for *cpa*, *cpb* and *cpx* genes

4. DISCUSSION

Many meals can be spoiled by anaerobic spore-forming, *Clostridium* species, and meat typically produces gas and/or foul odors (Kyrylenko et al., 2023). The development of *C. perfringens* outbreaks is known to be mostly influenced by improper chilling and holding temperatures, unsatisfactory food processing, and insufficient reheating and temperature management of meat (Leung et al., 2017). As can be observed in table (1), revealed more total anaerobic spore forming bacteria recovered from vacuum-packed salami were found by El-Rays (2014), who found that 28% of such microorganisms were associated from vacuum-packed salami, also anaerobic spore forming were more common in the vacuum-packed sausage samples than were examined in the samples reported by (Abd El- Rahman and WH 1996), who found that anaerobic bacteria were present in 80% of the examined samples. On the other hand, Ibrahim et al. (2015) found that the prevalence of anaerobic spore-forming bacteria isolated from meat products was 84%, 56%, and 0% for sausage, salami, and frankfurter, respectively. Shalout (1999), Mohammed (2002), El-Mossalami (2003), and Torky-Amal (2004) found that the mean values of the total anaerobic count of the sausage samples under examination were 2.62, 3.1, 5.59 and 5.22 log cfu/g, respectively. These results were differ from others as by Zakaria (2009) failed to detect anaerobic spore forming microorganisms in the examined vacuum-packed sausage samples and Ahmed et al. (2016) who isolated anaerobes from sausages with mean value  $1.5 \times 10^7 \pm 3.5 \times 10^6$  and according to the permissible limit stipulated by E.O.S (2005) for the total anaerobic count in salami (not more than 2 log CUF /g), 44% of the examined samples were accepted and 56% of the examined samples were unaccepted. Regarding to results of the bacteriological analysis of *Cl. perfringens* in the examined samples under

study table (4), The outcomes of Abd El Salam (2015) isolation of *Clostridium perfringens* from animal products were almost same, while these results were higher than that of El-Rays (2014) who detected anaerobic spore forming in salami and frankfurter with prevalence of 28% and 48%, respectively and Zakaria (2009) who failed to isolated anaerobic bacteria from sausage while the prevalence of anaerobic bacteria isolated from frankfurter was 13.3%. Similar results reported by Ibrahim et al. (2015) who reported that the prevalence of *Clostridium perfringens* in salami and sausage were 44% and 80%, respectively, while they failed to detect them in frankfurter. *Clostridium* spp. are frequently found in food and the environment so, they are a major cause of food poisoning and spoilage (Gribble et al. 2014). Since meat is not a sterile product, surface contamination with bacteria during processing is what causes post processing spoilage (Boerema et al., 2007). In addition, 16% and 28% of the examined samples of salami and frankfurter were contaminated with *C.perfringens*, respectively (El-Rays, 2014) and the count of *C.perfringens* were  $9.65 \times 10^2 \pm 2.08 \times 10^2$  and  $2.81 \times 10^3 \pm 0.44 \times 10^3$  for salami and frankfurter, respectively. Hamoda(2011) recorded that *C.perfringens* counts (cfu/g) from sausage were  $2.5 \times 10^3$ . Shalaby et al. (2011) examined 30 samples of sausage, the incidence of *C.perfringens* in the examined samples was 40% also, Ahmed et al. (2016) isolated *C.perfringens* from 33.3% of examined sausage. More poisons are produced by clostridial spp than by any other kinds of bacteria, *Clostridium perfringens* is the most common and greatest producer of toxins among the *Clostridium* species; it can be found in soil, animals' microbiota and human bodies. Due to their involvement in the etiology of *C. perfringens*-associated illness (CPAD) in both humans and animals, enterotoxin and beta toxin, two of the virulence components of *C. perfringens*, have drawn more attention than others (Monma et al., 2015). *C. perfringens* can be serotyped into five groups (A to E) based on the production of specific exotoxins [alpha ( $\alpha$ ), beta ( $\beta$ ), epsilon ( $\epsilon$ ), and iota ( $\iota$ )] (Moustafa et al., 2022). The  $\alpha$ -toxin, encoded by a plasmid mediated *cpa* gene, is associated with serotype A, as well as all other serotypes of *C. perfringens* (Cooper and Songer 2010). Meanwhile, the serotype B harbors *cpb* and *etx* plasmid-mediated genes that encode  $\beta$ - and  $\epsilon$ -toxins, respectively. The  $\beta$ - and  $\epsilon$ -toxins are associated with serotypes C and D, respectively (Chenet al., 2011), but serotype E has the plasmid-mediated *iap* gene, which produces  $\iota$ -toxin (Park et al. 2015). Christoph et al. (2004) recorded the toxin genes *cpa*, *cpb*, and *iap* using multiplex PCR; similar results were obtained by Bendary et al. (2022) and Ibrahim et al. (2015) as the authors found *cpa* from the analyzed materials. However, Deng et al. (2006) were unable to find *C. Perfringens cpb* gene. *Clostridium perfringens* is now the second most common bacterium linked to foodborne illness in the United States, accounting for one million cases annually, according to the Centers for Disease Control and Prevention (Grass et al. 2013).

## 5. CONCLUSIONS

From the obtained results it could be concluded that sausage samples were the most contaminated products. So, periodical examination and HACCP system application considered very important in meat product plants.

## CONFLICT OF INTEREST

The authors declare that they have no conflicts of interest for current data

## 6. REFERENCES

1. Abd El Salam, M.M. 2015. Bacteriological Attributes Of Vacuum Packed Meat Products Ph. D. Thesis, Fac. Vet. Med., Benha univ.
2. Abd El-Rahman, M. A., and WH, A. E. A. 1996. Anaerobic and aerobic aspects of bacterial contamination in frozen meat and meat products. Zagazig J. Pharm. Sci, 4, 227-240
3. Ahmed, I. I., Shaltout, F.A.a, Zakaria, I.M.b and Lamiaa, M. lotfyc ,2016. Bacterial evaluation of vacuum packed meat products. Benha Veterinary Medical Journal, 31,2 , 189-195.
4. APHA "American Public Health Association" 2001. Compendium of Methods for the Microbiological examination of Foods. 4th Ed. F.P. Downes and K. Ito ,editors , APHA. Washington D.C., USA.
5. Bendary, M. M., Abd El-Hamid, M. I., El-Tarabili, R. M., Hefny, A. A., Algendy, R. M., Elzohairy, N. A., and Moustafa, W. H. 2022. *Clostridium perfringens* associated with foodborne infections of animal origins: Insights into prevalence, antimicrobial resistance, toxin genes profiles, and toxinotypes. Biology, 11,4 , 551.
6. Boerema, J., Broda, D., Penney, N., Brightwell, G., 2007. Influence of peroxyacetic acid-based carcass rinse on the onset of "blown pack" spoilage in artificially inoculated vacuum-packed chilled beef. J. Food Protect. 70, 1434-1439.
7. Chang, S. H., Chen, C. H., and Tsai, G. J. 2020. Effects of chitosan on *Clostridium perfringens* and application in the preservation of pork sausage. *Marine Drugs*, 18,2 , 70-78
8. Chen, J., Rood, J. L., and McClane, B. A. 2011 Epsilon-toxin production by *Clostridium perfringens* type D strain CN3718 is dependent upon the agr operon but not the VirS/VirR two-component regulatory system. MBio, 2,6 , 10-1128.
9. Christoph G Baums, Ulrich Schotte, Gunter Amsberg, Ralph Goethe 2004. Diagnostic Multiplex PCR for toxin genotyping of *C. perfringens* isolates .Veterinary Microbiology, 100,1-2 , 11-16.
10. Collee, J.G.; Fraser, A.; Marmion, B. and Simmons, A. 1996 Practical Medical Microbiology. 14 Ed., Churchill Living Stone, New York, USA.
11. Cooper, K. K., and Songer, J. G. 2010. Virulence of *Clostridium perfringens* in an experimental model of poultry necrotic enteritis. *Veterinary microbiology*, 142,3-4 , 328-338.
12. Deng -Zhi ai, Li -XiaoQuan , Li -Chuanhua ,Zhag - Jian , and Duncan , C.L 2006. Recent developments of *C.perfringens* Food poisoning , 16<sup>th</sup> food Hygiene Symp. Teachers Food Hygiene Coll. Vet. Med. .94.
13. El-Mossalami, E. 2003. Risk assessment ready prepared meat products Ph. D.Meat Hygiene. Thesis, Fac.Vet.Med., Cairo Univ.
14. El-Rays,A. M.A. 2014 Aerobic and anaerobic spore formers in vacuum packed meat products Ph. D. Thesis, Fac. Vet. Med., Benha univ.
15. EOS ,Egyptian Organization for Standardization ,2005 . Meat products and their Quality Control, Arab Republic of Egypt.
16. Feldman, D., Ganon, J., Haffman, R. and Simpson, J. 2003. The solution for data analysis and presentation graphics. 2<sup>nd</sup> Ed., Abacus Lancrpts, Inc., Berkeley, USA.
17. Grass, J. E., Gould , L.H. and Mahon , B.E. 2013. Epidemiology of food borne disease outbreaks caused by *C. perfringens* Food borne pathogens and Disease.10,2 : 131-136.
18. Gribble, A., Mills, J., and Brightwell, G. 2014. The spoilage characteristics of *Brochothrixthermosphacta* and two psychrotolerant *Enterobacteriaceae* in vacuum packed lamb and the comparison between high and low pH cuts. Meat Science, 97,1 , 83-92.
19. Hamoda,W.S. 2011.study on the effect of some spices and organic salts on *C.perfringens* during of raw and cooked ground beef.Ph.D. Thesis, Fac. Vet.,Benha University.
20. Ibrahim, H. M., Amin, R. A., El-Shater, M. A., and Mohammed, M. M. 2015. Detection of *C. perfringens* toxins in vacuum packed meat products by using polymerase chain reaction. Benha Veterinary Medical Journal, 28,2 , 67-73.
21. ISO15213:2003 International Organization for Standardization, 2003b. Microbiology of food and animal feeding stuffs — Horizontal method for the enumeration of sulfite-reducing bacteria growing under anaerobic conditions.

22. Juneja, V. K., Gonzales-Barron, U., Butler, F., Yadav, A. S., and Friedman, M. 2013. Predictive thermal inactivation model for the combined effect of temperature, cinnamaldehyde and carvacrol on starvation-stressed multiple *Salmonella* serotypes in ground chicken. *International Journal of Food Microbiology*, 165,2 , 184-199.
23. Koneman, K., Allen, S., Dowell, V. and Sommers, H. 1992. *Color atlas and text book of diagnostic microbiology*. 2nd Ed., B. Lippincott Co., London, UK.
24. Kyrylenko, A., Eijlander, R. T., Alliney, G., Lucas-van de Bos, E., and Wells-Bennik, M. H. 2023. Levels and types of microbial contaminants in different plant-based ingredients used in dairy alternatives. *International Journal of Food Microbiology*, 110392.
25. Leung, V. H., Phan, Q., Costa, C. E., Nishimura, C., Pung, K., Horn, L., and Sosa, L. 2017. Notes from the Field: *Clostridium perfringens* outbreak at a catered lunch—Connecticut, September 2016. *Morbidity and Mortality Weekly Report*, 66,35 , 940-941.
26. Limbo, S., Torri, L., Sinelli, N., Franzetti, L., and Casiraghi, E. 2010. Evaluation and predictive modeling of shelf life of minced beef stored in high-oxygen modified atmosphere packaging at different temperatures. *Meat Science*, 84,1 , 129-136.
27. Meer, R.R. and Songer, J. G. 1997. Multiplex polymerase chain reaction assay for genotyping *Clostridium perfringens*. *Amer. J. Vet. Res.*, 58:702-705.
28. Mellou, K., Kyritsi, M., Chrysostomou, A., Sideroglou, T., Georgakopoulou, T., and Hadjichristodoulou, C. 2019. *Clostridium perfringens* foodborne outbreak during an athletic event in northern Greece, June 2019. *International Journal of Environmental Research and Public Health*, 16,20 , 3967- 3985
29. Mohammad, A. M., Chowdhury, T., Biswas, B., and Absar, N. 2018. Food poisoning and intoxication: A global leading concern for human health. In *Food safety and preservation* 307-352. Academic Press.
30. Mohammed, M.M.S. 2002. Quality improvement of some Egyptian frozen meat products Ph.D.Vet. Thesis, Meat Hygiene , Vet.Med., Cairo Univ
31. Monma, C, Hatakeyama, K, Obata, H, Yokoyama, K, Konishi, N, Itoh, T, Kai, A. 2015. Four foodborne disease outbreaks caused by a new type of enterotoxin-producing *Clostridium perfringens*. *J Clin Microbiol* 53:859 –867.
32. Moustafa, S., Zakaria, I., Moustafa, A., AboSakaya, R., & Selim, A. 2022 Molecular epidemiology and genetic characterization of *Clostridium perfringens* infections in lambs. *Microbial Pathogenesis*, 173, 105822.
33. Okeke, I. N., Laxminarayan, R., Bhutta, Z. A., Duse, A. G., Jenkins, P., O'Brien, T. F., and Klugman, K. P. 2005. Antimicrobial resistance in developing countries. Part I: recent trends and current status. *The Lancet Infectious Diseases*, 5,8 , 481-493.
34. Palmer, J., Flint, S., and Brooks, J. 2007. Bacterial cell attachment, the beginning of a biofilm. *Journal of Industrial Microbiology and Biotechnology*, 34,9 , 577-588.
- Limbo, S., Torri, L., Sinelli, N., Franzetti, L., and Casiraghi, E. 2010. Evaluation and predictive modeling of shelf life of minced beef stored in high-oxygen modified atmosphere packaging at different temperatures. *Meat Science*, 84,1 , 129-136.
35. Park, J. Y., Kim, S., Oh, J. Y., Kim, H. R., Jang, I., Lee, H. S., and Kwon, Y. K. 2015. Characterization of *Clostridium perfringens* isolates obtained from 2010 to 2012 from chickens with necrotic enteritis in Korea. *Poultry Science*, 94,6 , 1158-1164.
36. Ruiz-Capillas, C., and Herrero, A. M. 2019. Impact of biogenic amines on food quality and safety. *Foods*, 8,2 , 62.
37. Shaltout , F. A. 1999. Anaerobic bacteria in vacuum packed meat products .*BenhaVet .Med J.* , 10 ,1 : 1-10
38. Torky, A.A.Sh.A. 2004. Trails for inhibition of some food poisoning microorganism in meat products Ph.D. Thesis ,meat Hygiene , Fac. Vet. Med., Cairo Univ
39. Zakaria, I. M. 2009 *Clostridial* species and related anaerobic organisms in vacuum packed meat products with special reference to their shelf-life. Ph.D. Thesis, Benha University