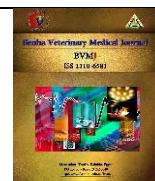




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Incidence of Enterobacteriaceae in some freshwater fishes

Rabab Rawash¹, Saad M. Saad¹, Fatin S. Hassanin¹, Mohamed A. Hassan¹, Maarouf A. Afifi²

¹ Department of Food Hygiene and Control, Faculty of Veterinary Medicine, Benha University, Egypt.

² Food Hygiene Department, Animal Health Research Institute, Benha Branch, Egypt

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ABSTRACT

It is evident that members of enterobacteriaceae are one of the most incriminated bacterial food poisoning causes. Salmonella serovars and pathogenic *E. coli* may be responsible for more than 15% of reported annual food poisoning cases; and with increasing price of beef and poultry meats, and consumer's fish meat desire and demand. So, this study aimed to spot the light on the prevalence of enterobacteriaceae members in some raw Nile fishes. 100 samples of *Oreochromis nilotica* (tilapia) and *Bagrus bayad* (Bayad), each of 50, were collected from different retail markets in Benha city, Qalubiya governorate, Egypt. Results revealed that 94% and 16% of examined samples were contaminated with enterobacteriaceae and coliform bacteria, with mean counts 2.65×10^3 and 1.81×10^3 CFU/g for tilapia samples, and 2.07×10^3 and 1.74×10^3 CFU/g for bayad samples, respectively. Bacteriological classification of enterobacteriaceae isolates revealed detection of *Citrobacter diversus*, *Citrobacter freundii*, *E. coli*, *Enterobacter aerogenes*, *Enterobacter cloacae*, *Klebsiella oxytoca*, *Proteus mirabilis*, *Proteus vulgaris*, *Yersinia enterocolitica* in variable percentages. 13% of examined samples were contaminated with *E. coli*, while 4% of examined tilapia samples only were contaminated with *Yersinia enterocolitica*. It is obvious that raw fishes may harbor bacterial contamination of significant health importance, so thorough cooking and processing hygienic practices are strongly recommended.

1. INTRODUCTION

Fish had long been regarded as a desirable and nutritional source of high-quality protein and generous supply of minerals and vitamins constituting the major part of human diet (Hastein *et al.*, 2006).

Tilapia nilotica and *Bagrus bayad* are two of the most widely distributed aquaculture commodities in Egypt (Sadek *et al.*, 2006). There has been a large increase in demand for tilapia and bayad, resulting in the increased production of both fishes through the method of high stocking density. However, the high stocking density method leads to rapid deterioration of water quality and increased fish stress (Sebastião *et al.*, 2015), resulting in bacterial invasion and proliferation in fish tissues.

Although, when a healthy fish is caught, the fish musculature is considered free of microbial contamination as its immune system prevents bacteria to proliferate easily whereas after death the immune system of fish collapses allowing easily penetration of microorganisms into the flesh (Kasing *et al.*, 1999). The penetration increase in case of fish caught from polluted area where there are high densities of bacteria (Sumner and Rose, 2002). Thus, many investigators convinced that fish from polluted environment may be passive carriers of pathogenic bacteria to man (Samaha *et*

al., 2004). In addition, contamination may be occurred during marketing at ground level or by hands (Sumner and Rose, 2002).

Enterobacteriaceae are a large, diverse heterogeneous group of rod-shaped Gram-negative bacilli that survive under aerobic conditions and normally inhabit the intestine of man and animals; some are motile while some others are not. The family includes many genera, some of which are part of the normal flora and incidentally cause diseases especially when given the opportunity. They are non-spore forming and some have capsules while others do not (Udeze *et al.*, 2012).

Enterobacteriaceae can be termed as common water-borne fish bacterial infections, which may normally in the tissues of apparently healthy fish, and the gastrointestinal tract of humans and animals (Newaj *et al.* 2008; Wogu and Maduakol, 2010), occurrence of fecal coliforms in fish was defined as a marker for the pollution level of the environment, they live in (CDC, 2013). Salmonella species and *E. coli* were the most repeatedly isolated (Sugumar *et al.*, 2008).

However, *Enterobacteriaceae* species including *Salmonella*, *E. coli* and *Yersinia enterocolitica* are not typical of water or of aquatic products as reported by FAO (2003), Aziz and Dapgh (2005), Pao *et al.* (2008) exemplified Salmonellae and

* Corresponding author: **Prof. Mohamed A. Hassan**. Department of Food Hygiene and Control, Faculty of Veterinary Medicine, Benha University, Egypt.

E. coli as a microorganism act as a public health hazard if present where they cause food poisoning.

Therefore, the present study was conducted to investigate the presence of *Enterobacteriaceae* species in retail *Tilapia nilotica* and *Bagrus bayad* fishes from Benha city markets.

2. MATERIAL AND METHODS

2.1. Collection of Samples:

A grand total of 100 random specimens of raw chilled freshwater fishes represented by *Tilapia nilotica* and *Bagrus bayad*, 50 of each, were collected randomly, along a year (March 2018 to February 2019), from retailers and supermarkets in Benha city, Qalyubia governorate, Egypt. The collected samples were kept frozen and transported to the lab where subjected to bacteriological examinations to evaluate the incidence of enterobacteriaceae contamination.

2.2. Preparation of samples according to (APHA, 2013)

Twenty-five grams of the sample were mixed with 225 ml sterile 0.1% peptone water. The contents were homogenized at Stomacher (M A 106402, France, 450 to 640 strokes per minute) for 2 minutes, and 1 ml of the mixture was transferred into separate tube each containing 9 ml sterile 0.1% peptone water, from which tenth-fold serial dilutions were prepared. The prepared samples were subjected to the following bacteriological examination:

2.3. Total Enterobacteriaceae count (ICMSF, 1996)

0.1 ml from each of the previously prepared dilution was spread on Violet Red Bile Glucose agar (VRBG) and incubated at 37 °C for 24 hours. All purple colonies were counted, and the average number of colonies was determined.

2.3.1. Identification of family Enterobacteriaceae

It was conducted according to Cowan and Steel (1974) performed by Gram's stain, Biochemical tests, and motility tests.

2.4. Determination of coliform count (ISO 4832, 2006).

One ml from each of the previously prepared dilution was cultured in Violet Red Bile agar (VRBA) by pour-plate technique and incubated at 37 °C for 24 hours. All purple colonies were counted, and the average number of colonies was determined.

2.5. Isolation and identification of *E. coli*

2.5.1. Isolation of *E. coli* according to (ISO 16649-2, 2001).

1 ml from each of the previously prepared dilution was cultured in Tryptone-Bile-Glucuronic Agar (TBX) by pour-plate technique and incubated at 44 °C for 24 hours. All bluish-green colonies were counted, and the average number of colonies was determined.

2.5.2. Identification of *E. coli*.

2.5.2.1. Gram's Stain according to (Cruickshank *et al.*, 1975).

2.5.2.2. Biochemical tests (MacFaddin, 2000).

2.5.2.3. Serological Identification according to Kok *et al.* (1996).

2.6. Isolation and identification of *Salmonellae*

2.6.1. Isolation of salmonellae according to (ISO 6579, 2017).

2.6.1.1. Pre-enrichment in non-selective buffered peptone water broth, which then incubated at 37±1 °C for 18±2 hours.

2.6.1.2. Enrichment in Rappaport Vassiliadis broth (RV broth), then the tube was incubated at 43°C for 24 hours.

2.6.1.3. Selective Plating on Xylose lysine Desoxy chocolate (XLD) agar and Brilliant Green agar. The plates were incubated at 37°C for 24 hours. Plates were examined for suspected *Salmonella* colonies which appeared as red with black centers on XLD agar and pink on Brilliant Green agar.

2.6.2. Identification of salmonellae

2.6.2.1. Gram's Stain (Cruickshank *et al.*, 1975).

2.6.2.2. Biochemical identification (Krieg and Holt, 1984).

2.6.2.3. Serological identification (Confirmatory test) according to (Kauffmann, 1974).

2.7. *Statistical analysis*: of the obtained results was performed according to Feldman *et al.* (2003).

3. RESULTS

Results demonstrated in table (1) showed that *Enterobacteriaceae* were equally detected at rate of 94% of the examined tilapia and bayad samples, with mean values 2.65×10^3 and 2.07×10^3 cfu/g, respectively. The statistical results revealed that, there was a significant ($P \leq 0.05$) increase of *Enterobacteriaceae* counts of Nile tilapia when compared with Bayad samples.

Following results summarized in table (2) showed that coliform bacteria were detected in the examined tilapia and bayad samples with incidence of 14 and 18%, with mean values 1.81×10^3 and 1.74×10^3 cfu/g, respectively. The statistical results revealed that, there is no significant difference between tilapia and bayad samples.

Table (3) demonstrated a detailed bacteriological classification of *Enterobacteriaceae* isolates which revealed detection of *Citrobacter diversus*, *Citrobacter freundii*, *E. coli*, *Enterobacter aerogenes*, *Enterobacter cloacae*, *Klebsiella Oxytoca*, *Proteus mirabilis*, *Proteus vulgaris*, and *Yersinia enterocolitica* in different ratios. On the other hand, *Salmonella* sp. not detected in any of the examined tilapia and bayad samples.

Table (4) showed the serotyping of isolated *E. coli* strains revealed detection of O27:H2, O63:H2, and O158:H4 with incidences of 8, 2, 4% and 6, 2, 4% for tilapia and bayad samples, respectively.

Moreover, following (EEC, 2005) specifications, 14% and 4% of the examined tilapia samples were rejected due to detection of *E. coli* and *Y. enterocolitica*; while 12% of bayad samples were rejected due to detection of *E. coli* as shown in table (5).

Table 1 Total Enterobacteriaceae counts cfu/g in the examined samples of Nile fish (n=50).

Types of Samples	Positive		Min.	Max.	Mean ±S.E.
	No.	%			
Nile tilapia	47	94.0	3.0×10^2	4.90×10^3	$2.65 \times 10^3 \pm 0.16 \times 10^3^a$
Bayad	47	94.0	3.0×10^2	4.50×10^3	$2.07 \times 10^3 \pm 0.01 \times 10^3^b$
Total*	94	94.0			

^{a, b} Mean value within a column followed by difference letter where significant difference ($P \leq 0.05$). Total*: number and percentage in relation to total number of samples (100).

Table 2 Total Coliform counts (cfu/g) in the examined samples of Nile fish (n=50).

Types of Samples	Negative		Positive		Min.	Max.	Mean \pm S.E.
	No.	%	No.	%			
Nile tilapia	7	14.0	43	86.0	3.0×10^2	4.00×10^3	$1.81 \times 10^3 \pm 0.13 \times 10^{3a}$
Bayad	9	18.0	41	82.0	1.0×10^2	2.90×10^3	$1.74 \times 10^3 \pm 0.10 \times 10^{32a}$
Total*	16	16.0					

^{a, b}: Mean value within a column followed by difference letter where significant difference ($p \leq 0.05$). Total*: number and percentage in relation to total number of samples (100).

Table 3 Prevalence of *Enterobacteriaceae* species in examined samples of Nile fish (n=50).

Bacterial isolates	Samples				Total No. & %**
	Nile tilapia (<i>O. niloticus</i>)		Bayad (<i>B. bayad</i>)		
	No.	%*	No.	%*	
<i>Citrobacter diversus</i>	2	4.0	1	2.0	3
<i>Citrobacter freundii</i>	1	2.0	1	2.0	2
<i>Edwardsiella tarda</i>	0	0.0	0	0.0	0
<i>E. coli</i>	7	14.0	6	12.0	13
<i>Enterobacter aerogenes</i>	1	2.0	1	2.0	2
<i>Enterobacter cloacae</i>	1	2.0	1	2.0	2
<i>Kl. pneumoniae</i>	0	0.0	0	0.0	0
<i>Kl. oxytoca</i>	3	6.0	3	6.0	6
<i>Proteus mirabilis</i>	1	2.0	1	2.0	2
<i>Proteus vulgaris</i>	1	2.0	1	2.0	2
Salmonella spp.	0	0.0	0	0.0	0
<i>Y. enterocolitica</i>	2	4.0	0	0.0	2
Total	19	38.0	15	30.0	24

%*: Percentage in relation to total number of each sample (50). %***: Percentage in relation to total number of samples (100).

Table 4 Serotyping of *E. coli* isolated from examined samples of Nile fish (n=50)

<i>E. coli</i> strains	Samples				Total No. & %**	Strain characteristic
	Nile tilapia (<i>O. niloticus</i>)		Bayad (<i>B. bayad</i>)			
	No.	%*	No.	%*		
O27:H2	4	8.0	3	6.0	7	EPEC
O63:H2	1	2.0	1	2.0	2	EPEC
O158:H4	2	4.0	2	4.0	4	EPEC
O159:H7	0	0.0	0	0.0	0	EHEC
Total	7	14.0	6	12.0	13	-

%*: Percentage in relation to total number of each sample (50). %***: Percentage in relation to total number of samples (100). EPEC: Enteropathogenic *E. coli*. ETEC: Enterotoxigenic *E. coli*. EHEC: Enterohaemorrhagic *E. coli*

Table 5 Incidence and acceptability of *E. coli* in the examined samples of Nile fish (n=50).

Microorganism	Fish samples							
	Nile tilapia (<i>O. niloticus</i>)				Bayad (<i>B. bayad</i>)			
	rejected		accepted		rejected		accepted	
No.	%*	No.	%*	No.	%*	No.	%*	
<i>E. coli</i>	7	14.0	43	86	6	12.0	44	88
<i>Y. enterocolitica</i>	2	4.0	48	96	0	0.0	50	100
<i>Salmonella</i> Sp.	0	0.0	50	100	0	0.0	50	100

%*: Percentage was recorded according to number of each sample (50).

4. DISCUSSION

Enterobacteriaceae especially food poisoning members represent a major and important group of microorganisms because of their frequent occurrence and activities that may

have a negative impact on seafood quality. The presence of human pathogenic bacteria in fish and seafood may be attributed to contamination during processing. Several bacteria are, however, reported to cause infection and mortality in both fish and humans (Hastein *et al.*, 2006).

Results illustrated in table (1) showed the incidence and counts of *Enterobacteriaceae* in the examined samples which were nearly similar to those reported by Engelbrecht *et al.* (2008) and El-Shabasy (2009) (3.8×10^3 and 2.8×10^3 cfu/g for tilapia and bayad samples, respectively). Lower results were recorded by Gaafar (2007) (7.8×10^5 and 6.24×10^5 cfu/g for tilapia and bayad samples, respectively). On the other hand, the present results were higher than those recorded by El-Sherief (2015) (7.64×10^2 cfu/g in examined tilapia samples), and Hassan (2013) (2.9×10^4 cfu/g for bayad samples).

Following results of coliform tabulated in table (2), results were in agree with Engelbrecht *et al.* (2008) and El-Shabasy (2009) (0.98×10^3 and 2.4×10^3 cfu/g for tilapia and bayad samples, respectively), while were lower than those recorded by Gaafar (2007) (2.9×10^5 and 1.1×10^5 CFU/g for tilapia and bayad samples, respectively), also, they were higher than those recorded by Naser (1991) (5.8×10^2 cfu/g for tilapia samples).

Results of bacteriological classification of the isolated *Enterobacteriaceae* isolates as summarized in table (3) were in agreement with those recorded by Abo Samra (2001) who isolated *E. coli*, *Klebsiella oxytoca*, *Enterobacter cloacae*, *Enterobacter aerogenes*, *Citrobacter freundii*, *Proteus mirabilis*, *Proteus vulgaris* from examined freshwater fishes' samples).

The incidences of *E. coli* were nearly similar to those recorded by El-Sherief (2015) and Aki (2009) (12% and 16% for tilapia and bayad samples), while were lower than those recorded by Hassan (2013) (57.1 and 100% of examined tilapia and bayad samples, respectively), also, they were higher than those recorded by Ibrahim (2014) and Gaafar (2007) (8% for both tilapia and bayad samples).

The detected values of *Y. enterocolitica* detection in tilapia samples were nearly similar to these recorded by Aki (2009) (4%), while were lower than those recorded by Ammar (2016) (20%). The failure in detection of *Y. enterocolitica* from bayad samples is in agree with Aki (2009).

In addition, the failure of *Salmonella* detection in both tilapia and bayad samples is in agree with Papadopoulou (2007), while disagreed with Hassan (2013) and Ibrahim (2014) who detected *Salmonella* in examined samples at rate of 57.1 and 8%, respectively.

Variations between authors may be attributed to the differences in pollution levels of the environment of fish rearing, mishandling and improper storage practices, and the immunity status of the fish.

5. CONCLUSION

It could be concluded that the presence of *Enterobacteriaceae*, coliform and their members indicating several faults and improper practices of rearing, mishandling and improper storage properties during catching and marketing, that highlighted low hygienic knowledge and encourage authorities for more restrictions and observations, and recommend thorough heat treatment before consumption.

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CONFLICT OF INTREST

The content of this report solely reflects the opinions of the authors, and we report no conflicts of interest. FAPESB did not play a role in the research or writing of the paper.

6. REFERENCES

1. Abo Samra, R.G. 2001. Hygienic evaluation of some fishes marketed in Damietta. Thesis, Master of Veterinary Medicine (Meat Hygiene Dept.), Zagazig University, Egypt.
2. Aki, M.A.R. 2009. *Enterobacteriaceae* in fresh water fish. Thesis, Ph.D. of Veterinary Medicine (Meat Hygiene Dept.), Alexandria University, Egypt.
3. American Public Health Association "APHA", 2013. Compendium of methods for the microbiological examination of food. T. Matthew Taylor, John N. Sofos, Peter Bodnaruk, and Gary R. Acuff (Eds.), 4th Ed., Ch. 2, Washington DC., USA.
4. Ammar, S.A. 2016. Advanced studies on enteric pathogens of fresh water fish in Kafr El-Sheikh province. Thesis, Ph.D. of Veterinary Medicine (Meat Hygiene Dept.), Alexandria University, Egypt.
5. Aziz, H., Daphn, A. 2005. Bacteriological studies of fecal and water samples from different sources with special reference to some Gram-negative bacteria. Benha Vet. Med. J., 16 (2): 248-261.
6. Centers for Disease Control and Prevention (CDC). 2013. Surveillance for foodborne disease outbreaks-United States, 2009-2010. *Morb. Mortal. Wkly. Rep.*, 62(3):41-47.
7. Cowan, S.T., Steel, K.J. 1974. Manual for identification of medical bacteria. Cambridge Univ. Press, London, New York, Malburne.
8. Cruickshank, R., Duguid, J.P., Marmion, R.H., Swain, R.H. 1975. *Medical Microbiology*. 12th Ed., Edinburg, London and New York.
9. EEC, 2005. Commission regulation (EC) No.2073/2005 on microbiological criteria for foodstuffs. Council of the European Communities (EEC). *Off. J. Eur. Commu.* 1.338:22.
10. El-Shabasy, N.A.A. 2009. Microbiological analysis of farm fishes in Alexandria governorate. Thesis, Ph.D. of Veterinary Medicine (Meat Hygiene Dept.), Alexandria University, Egypt.
11. El-Sherief, M.F.A. 2015. *Enterobacteriaceae* associated with farm fish and retailed ones. Thesis, Master of Veterinary Medicine (Meat Hygiene Dept.), Alexandria University, Egypt.
12. Engelbrecht, K., Silva, J., Daton, A. 2008. Microbial evaluation of fresh aquacultured fish. *J. Applied Bacteriology*, 49(8): 274-278.
13. Feldman, D., Ganon, J., Haffman, R., Simpson, J., 2003. The solution for data analysis and presentation graphics. 2nd Ed., Abacus Lancripts, Inc., Berkeley, USA.
14. Food and Agriculture Organization (FAO) 2003. Assessment and management of seafood safety and quality. FAO fisheries technical paper 444.
15. Gaafar, H.M. 2007. Microbiological quality of retail fresh fish in Cairo. Thesis, Master of Veterinary Medicine (Meat Hygiene Dept.), Alexandria University, Egypt.
16. Hassan, H.R.M. 2013. Enteropathogens in some freshwater fishes. Thesis, Master of Veterinary Medicine (Meat Hygiene Dept.), Alexandria University, Egypt.
17. Hastein, T., Hjeltne, B., Lillehaug, A., UtneSkåre, J., Berntssen, M., Lundebye, A.K. 2006. Food safety hazards that occur during the production stage: challenges for fish farming and the fishing industry. *Rev. Sci. Technol.*, 25(2): 607-625.
18. Howgate, P. 1985. The shelf-life of fish products. *J. Sci. Food Agric.*, 36(2) 12: 126-127.
19. Ibrahim, S.Y.A. 2014. Prevalence of some food poisoning organisms in fish. Thesis, Master of Veterinary Medicine (Meat Hygiene Dept.), Zagazig University, Egypt.
20. International commission of Microbiological Specification for Foods "ICMSF" 1996. *Microorganisms in Food. I-Their Significance and methods of enumeration*. 3rd Ed., Univ. of Toronto, Canada.
21. International Organization for Standardization "ISO" 2001. International Organization for Standardization. No.16649-2. Microbiology of food and animal feeding stuffs-Horizontal method for the enumeration of glucuronidase-positive *Escherichia coli* - Part 2: Colony-count technique at 44 °C using 5-bromo-4-chloro-3-indolyl-D-glucuronide.
22. International Organization for Standardization "ISO" 2006. International Organization for Standardization. No.4832/2006. Microbiology of food and animal feeding stuffs-horizontal method for the enumeration of coliforms: colony count technique.
23. International Organization for Standardization "ISO" 2017. International Organization for Standardization. No.6579-1. Microbiology of the food chain Horizontal method for the detection, enumeration and serotyping of Salmonella - Part1: Detection of *Salmonella* spp.
24. Kasing, A., Asiah, M.Y., Kumbang, J. 1999. Distribution of bacteria in tropical freshwater fish and ponds. *International Journal of Environmental Health Research*, 9: 285-292.
25. Kauffmann, G. 1974. Kauffmann white scheme. WHO, BD 172, L. Rev. 1. *Acta Pathologica et Microbiologica Scandinavica*, 61: 385.
26. Kok, T., Worswich, D., Gowans, E. 1996. Some serological techniques for microbial and viral infections. In: *Practical Medical Microbiology*, Collee, J., Fraser, A., Marmion, B. and Simmons, A. (Eds.), 14th Ed., Edinburgh, Churchill Livingstone, UK.
27. Krieg, N.R., Holt, J.G. 1984. *Bergey's manual of systemic bacteriology*, Vol.1, William and Wilkins, Baltimore, M.D.21202, USA.
28. MacFaddin, J.F. 2000. *Biochemical tests for identification medical bacteria*. 3rd Ed., Williams and Wilkins, Philadelphia, P.A., USA.
29. Naser, G.N. 1991. Occurrence of some food poisoning microorganisms in natural and farm fishes. Thesis, Master of Veterinary Medicine (Meat Hygiene Dept.), Alexandria University, Egypt.
30. Newaj, A., Mutani, A., Ramsbhag, A., Adesiyun, A. 2008. Prevalence of bacterial pathogens and their anti-microbial resistance in Tilapia and their pond water in Trinidad. *Zoonosis Public Health*, 55(4): 206-213.
31. Pao, C., Molla, B., Kleer, J., Reine, A. 2008. Hygienic control of fish processing plant. *Wochenschr*, 121(4): 89-93.
32. Papadopoulou, C., Economou, E., Zakas, G., Salamoura, C., Dontorou, C., Apostolou, J. 2007. Microbiological and pathogenic contaminants of seafood in Greece, *J. Food Quality*, 30: 28-42.
33. Sadek, S., Osman, M. F., Mezayen, A. 2006. Aquaculture in Egypt: A Fragile Colossus?. AQUA 2006 International Conference and Exhibition, Firenze (Florence, Italy, May 9-13, 2006).
34. Samaha, H., Hassanien, R., Suwsan, A. 2004. Listeriosis in fish and abortion cases in women. *Minufiya Vet. J.*, 3 (1): 93-98.
35. Sebastião, F.A., Furlan, L.R., Hashimoto, D.T., Pilarski, F. 2015. Identification of bacterial fish pathogens in Brazil by

- direct colony PCR and 16s rRNA gene sequencing. Adv. Microbiol., 5(6): 409-424.
36. Sugumar, G., Chrisolite, B., Velayutham, P., Selvan, A., Ramesh, U. 2008. Occurrence and seasonal variation of bacterial indicators of faecal pollution along Thoothukudi Coast, Tamil Nadu. J. Environ. Biol., (3):387-91.
 37. Sumner, W. and Rose, S. 2002. Occurrence of potential pathogens in fish at retail level. J. Environ. Health Res., 12(3): 268 - 273.
 38. Udeze, A.O., Talatu, M., Ezediokpu, M.N., Nwanze, J.C., Onoh. C., Okonko. I.O. 2012. The effect of *Klebsiella pneumoniae* on catfish (*Clarias gariepinus*). Researcher, 4(4): 51-59.
 39. Wogu, M.D., Maduakol, M. 2010. Evaluation of microbial spoilage of some aqua cultured fresh fish in Benin City Nigeria. Ethiopian Journal of Environmental Studies and Management, 3(3):18-22.